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Uroderma bilobatum. By Robert J. Baker and Cora L. Clark

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Uroderma bilobatum Peters, 1866
Peters' Tent-making Bat

Uroderma bilobatum Peters, 1866:394. Type locality Sao Paulo, Brazil.
Uroderma convexum Lyon, 1902:83. Type locality Colon, Panama.

CONTEXT AND CONTEXT: Order Chiroptera, family Ptilostomidae, subfamily Stenodermatinae. The genus Uroderma has two species (Davis, 1968), Uroderma bilobatum and U. magnirostrum. Six subspecies of bilobatum are recognized (Baker and McDaniel, 1972; Davis, 1968) as follows:

U. b. bilobatum Peters, 1866:394. See above.
U. b. trinitatatum Davis, 1968:690. Type locality Caroni County, Trinidad.
U. b. thomasi Andersen, 1908:390. Type locality Bellavista, Departamento de Beni, Bolivia.
U. b. convexum Lyon, 1902:83. See above.
U. m. molaris Davis, 1968:696. Type locality Puleneque, Chiapas, Mexico.

DIAGNOSIS. Uroderma bilobatum has bilobed upper medial incisors; a narrow white line on middle of upper back; a distinct white line above and below each eye (Fig. 1a); no external tail; hairless posterior border of uropatagium; dorsal profile of skull from crown to tip of snout usually markedly depressed in frontal region (Fig. 2); mesethmoid much less expanded laterally; and facial stripes more pronounced (Davis, 1968) in comparison with U. magnirostrum.

GENERAL CHARACTERS. Geographic variation in Uroderma bilobatum was reviewed by Davis (1968) who recognized five subspecies based on cranial and pelage color variation. A sixth subspecies subsequently was described (Baker and McDaniel, 1972) based on chromosomal characters.

The skull (Fig. 2) and dentition of U. bilobatum was described by Miller (1907). The dental formula is 1/2, c/1, p/2, m 3/3, total 32. The published records of cranial and forearm measurements were reviewed by Swanepoel and Genoways (1979). Selected average external and cranial measurements (Swanepoel and Genoways, 1979) for eight specimens (four males and four females) from Trinidad and Nicaragua are (in mm, extremes in parentheses): length of forearm, 41.7 (39.6 to 43.1), greatest length of skull, 23.5 (22.6 to 24.7), condylobasal length, 21.0 (19.9 to 21.6), zygomatic breadth, 12.9 (12.5 to 13.4), postorbital constriction, 5.4 (4.6 to 5.7), breadth of braincase, 9.6 (9.3 to 9.9), length of maxillary toothrow, 8.1 (7.8 to 8.6), breadth across upper molars, 9.4 (8.9 to 9.9).

The nose leaf is simple but well developed; the ears are rimmed with pale yellowish pigment; the eyes are large (Fig. 1a); a calcare is present; and the dactylopsagius minus is broad, slightly pigmented and permanently open (Miller, 1907).

DISTRIBUTION. Uroderma bilobatum (Figs. 3, 4) is known from the lowlands of tropical America from Oaxaca and Veracruz, Mexico, south to Peru, Bolivia, and southeastern Brazil (Davis, 1968). Most records are from below 600 m elevation, but specimens have been taken in Central America as high as 1,500 m, and the altitudinal record for South America is 1,800 m at Guayabamba, Peru (Davis, 1968).

FORM AND FUNCTION. Secondary sexual dimorphism was noted (Baker et al., 1972) in samples of U. b. davisi (2 of 25 characters: zygomatic breadth and breadth across canines) and U. b. molaris (8 of 25 characters: skull length, condylobasal length, mandible length, occipitonasal length, depth of brain case, palatal length, zygorostral length, and breadth across canines). In all instances, males were larger than females.

The stomach and gut anatomy of U. bilobatum shares a suite of features characteristic of stenodermatines in general (Forman et al., 1979). The stomach of Uroderma is intermediate in form between Sturira and Artibeus. The caecum is long and has a narrow apex (Forman et al., 1979). The anatomy of the gastric muscosa in Uroderma is different from that of Vampyressa, Vampyrops, Chiroderma, and Sturira in that in Uroderma rugae are distributed diagonally only within the mid-region of the stomach, whereas in the other stenodermatine genera examined, these rugae are restricted to the caecum (Forman et al., 1979). Sperm morphology of U. bilobatum is similar to that of Artibeus jamaicensis (Forman and Genoways, 1979). The sperm head of U. bilobatum has an unusually short acrosome that covers an extremely small portion of the nucleus. The flattened base of the head with its pointed corners is unique among the stenodermatines examined (Forman and Genoways, 1979).

Internal and external brain anatomy of U. bilobatum is characteristic of stenodermatine species (McDaniel, 1976). The brain has a deep cerebrum with large pseudotemporal lobes that project ventrally but not to the extent observed in Artibeus. The sulci anterior to the pseudocentral sulci and the pseudocentral sulci are well developed. The inferior colliculi are not exposed dorsally and the vermilliform body of the cerebellum forms a medial crest. At the

Fig. 1. Tent constructed by Uroderma bilobatum from a banana leaf; (a) tent occupied by 11 individuals, (b) line (between arrows) where bats have chewed veins in the leaf to make a more protective tent. Photographs taken at Cana, Panama by R. J. Baker.
and pregnant have been taken. There is some evidence that females have a postpartum estrus (Fleming et al., 1972). In Panama, parturition was correlated with the wet and dry seasons and time of fruiting of plants. In samples from Guatemala, El Salvador, Honduras, and Nicaragua taken during the same year, there seemed to be a geographic variation in the timing of the reproductive cycles (Baker, 1981). In samples studied by Baker (1981), females were in relatively early stages of pregnancy in May and early June indicating that samples from northern Central America are not synchronized with those from Panama (Fleming et al., 1972). However, samples of 122 females from the Pacific versant of Central America support the conclusions of Fleming et al. (1972) that local populations are synchronized seasonally (Baker, 1981). Young bats become independent from females at about 1 month of age (Fleming et al., 1972).

ECOLOGY. Uroderma bilobatum is a fruit-eating bat with a local distribution strongly influenced by availability of tall-growing tropical vegetation (Baker, 1981; Greenbaum, 1981). Gardner (1977) concluded that U. bilobatum primarily is frugivorous, but also may feed on pollen, nectar, and insects associated with flowers and fruit. Most reports of food habits for the species simply note that this bat eats fruit (Goodwin, 1946; Tamsitt and Valdivieso, 1965; Villa-R., 1967). Biodell (1955) reported U. bilobatum to eat the pericarp of small unidentified palm fruits in Panama, de Carvalho (1961) recorded fruits of three species of Ficus (Ficus) as food items of the species in Brazil, and Goodwin and Greenhall (1961) found remains of Psidium guajava in two U. bilobatum in Trinidad. An analysis of contents of 320 stomachs of U. bilobatum collected from Costa Rica and Panama indicated a diet (percent of volume) of 76% plant material (unidentified), 13% insects, and 11% unclassified material (Fleming et al., 1972).

Endoparasites recorded from U. bilobatum include Trypanosoma cruzi and a Trypanosoma cruzi-like protozoan (Ubelaker et al., 1977). Ectoparasites include Amblyomma sp., Basilis constrieta, B. myotis, Chirotherobechia urdeanae, Entomobius latipes, Macronyssoides sp., Neotrichobius delicatus, Ornithodorus hasei, Paraseceia manueli, Parabatrachus dunn, P. longicuris, Periglauca therigi, Phylostomomyos anodryanerki, Trichobius costalimai, T. joblingi, T. keenani, and T. urodermae (Webb and Loenin, 1977). Uroderma bilobatum occurs over a wide geographic range and a list of species that have been taken in mist nets with U. bilobatum is extremely long and, at this time, virtually nondefinitive. The long list of associated species does indicate diversity in the ecology of U. bilobatum.

BEHAVIOR. Uroderma bilobatum rests singly or in groups ranging from 2 to 50 (Kunz, 1982). The most striking aspect of the biology of the species is the self-directed behavior of colonies to produce "tents" (Fig. 1a, b) by modifying leaves (Barbour, 1932; Kunz, 1982). Tents are constructed by weakening or severing the structural veins of leaves by biting or chewing. Foster and Tunn...
bilibatum was compared to those of representatives of Artibesia, Starnira, and Enchisthenes (Baker et al., 1979). Although the diploid number and fundamental number differ between the two groups (2n=44 in Uroderma but 30 to 31 in the other three genera), most of the euchromatic linkage groups could be identified among the genera. Also, C-band material was restricted to the centromeric region in all karyotypes studied.

ETYMOLOGY. The word *Uroderma* is from the Greek words *uro*, meaning tail, and *derma*, skin, referring to the presence of a uropatagium without tail vertebrae in this species. The species name *bilibatum* combines the prefix *bili* (Greek and Latin) meaning two, with *lobat*—(Greek and Latin) for lobed. This name describes the bilobed condition of the first upper incisors.

LITERATURE CITED


**FIG. 4.** Map showing the distribution of *Uroderma bilobatum* in South America. (1) *U. b. conexus*, (2) *U. b. bilobatum*, (3) *U. b. thomasi*, (4) *U. b. trinitatum*. (Map modified from Davis, 1968.)

(1976) reported that the number of bites required to sever the veins of a frond ranged from 44 in simple bilobed leaves of *Asterogyne*, to 80 in the large leaves of *Prickardia*. Tents constructed from *Prickardia paracribera*, *Cocos nucifera*, *Liriope chinenica*, *Sabal maauleiformis*, and *Maia* have been observed (Barbour, 1932; Foster and Timm, 1976). Details on the age at which bats make tents, the degree of cooperation between individuals, and whether this behavior is innate or learned are unknown (Kunz, 1982). Little is known about the activity patterns of *U. bilobatum*. Eckert (1982) suggested that time of capture of *Uroderma* in nets indicates a bimodal activity pattern. When roosting in tents by day, individuals are alert and fly immediately upon being disturbed.

GENETICS. *Uroderma bilobatum* has at least three chromosomal races (Baker and Lopez, 1970) which have been studied by G- and C-bandling. The 2n=38 and 2n=44 chromosome races are derived from a telomere-centromere translocation, involving two acrocentrics, a telomere-centromere translocation involving a small biarmed element and an acrocentric and a centric fusion (Baker, 1979). The 2n=44 and the 2n=42 chromosome races are distinguished by a centric fusion and a pericentric inversion (Baker et al., 1982).

The contact zone between the 2n=44 and the 2n=38 chromosomal races is located on the Pacific versant of Middle America in El Salvador, Honduras, and Nicaragua. This zone has been the object of intensive chromosomal (Baker, 1979, 1981; Baker et al., 1979) and electrophoretic (Greenbaum, 1981) studies. Backcross individuals have been found over a wide range (more than 400 km) but F1's are known over a much smaller distance (less than 50 km). Populations at the center of the zone are composed mostly if not entirely of hybrid individuals (Baker, 1981). Electrophoretically, the two chromosomal races are similar (genetic similarity for all populations was greater than 0.96; Greenbaum, 1981). The evolutionary implications of data from the *Uroderma* hybrid zone have been debated extensively (Barton, 1982; Hafner, 1982). Baker (1981) emphasized that the biology of *Uroderma* does not fit the parameters (low vagility, high prolificity, and highly inbred demes) most often described as characteristic of species that undergo extensive chromosomal evolution. He further concluded that the data from *Uroderma* indicate that the negative heterosis assumption in models of chromosomal evolution is often an overstatement. Barton (1982) disagreed with Baker (1981) and Greenbaum (1981) and concluded that the chromosomal contact zone in *Uroderma* shows all the characteristics expected from a hybrid zone maintained by moderate selection against heterozygotes or recombinant genotypes, whereas Hafner (1982) concluded that the zone does not depart significantly from that predicted by a model of simple diffusion of one parental form into another parental form. The positions of Barton (1982) and Hafner (1982) are not compatible and at least one position is incorrect.

The G- and C-band karyotype of the 2n=44 cytotype of *U.
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