

# Physical requirements

- Temperature
- Oxygen
- pH
- Osmotic conditions

# Temperature

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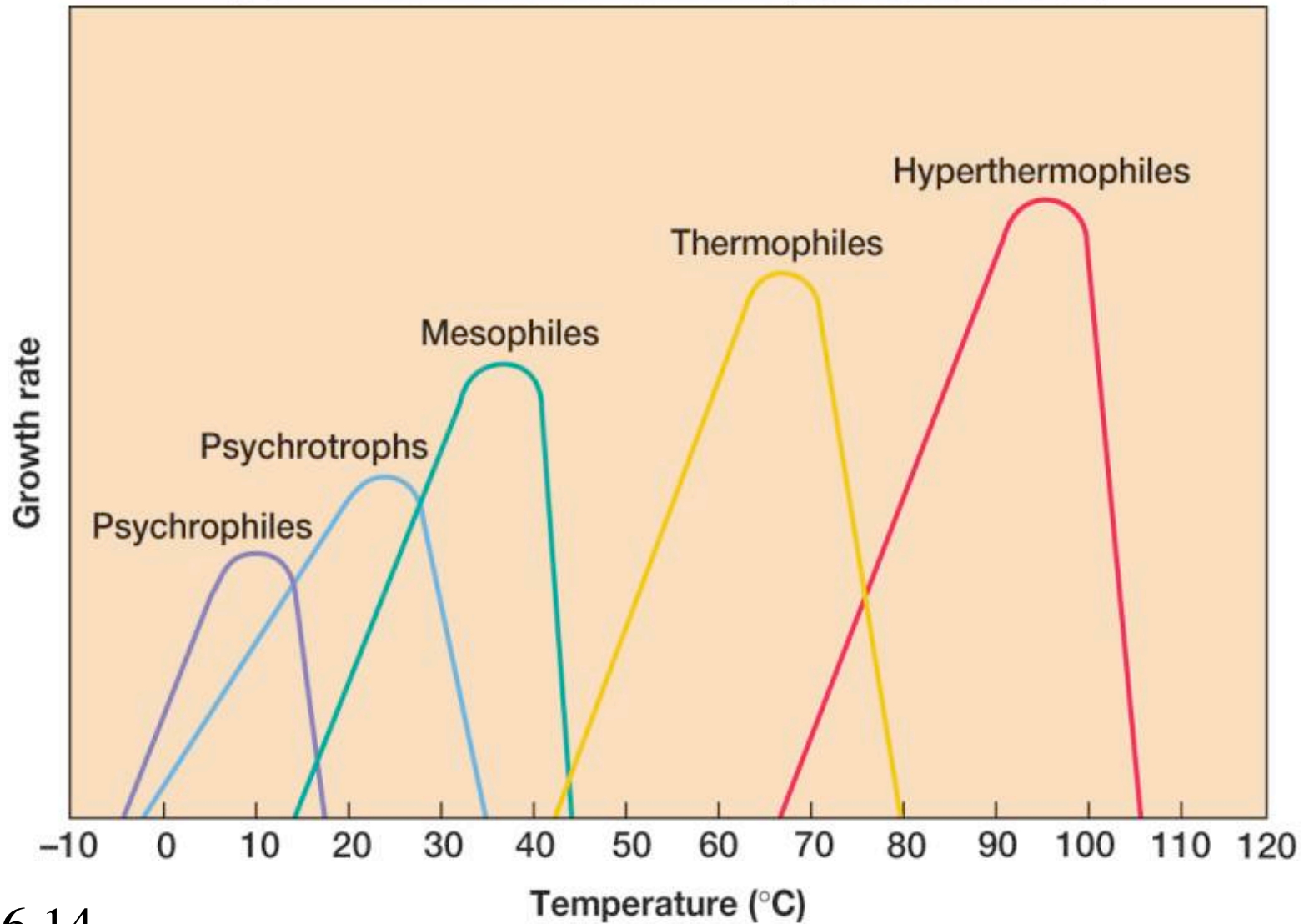
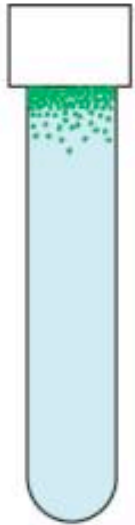


Fig. 6.14

# Oxygen

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Obligate  
aerobe



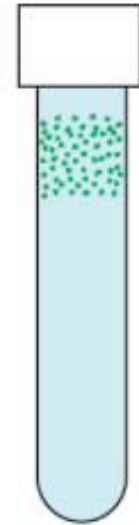
Facultative  
anaerobe



Aerotolerant  
anaerobe



Strict  
anaerobe



Microaerophile

## Enzyme content

+ SOD  
+ Catalase

+ SOD  
+ Catalase

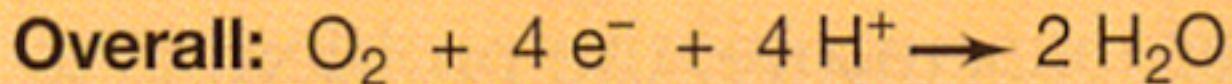
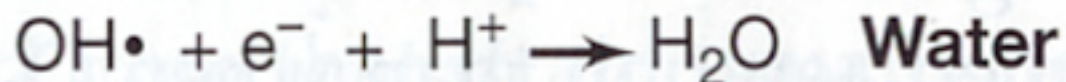
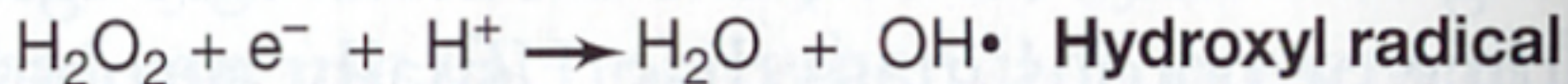
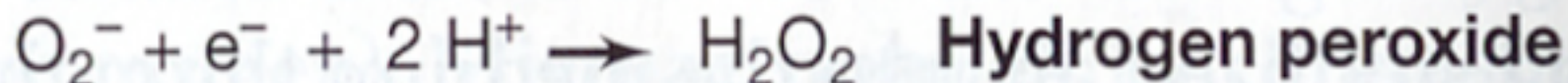
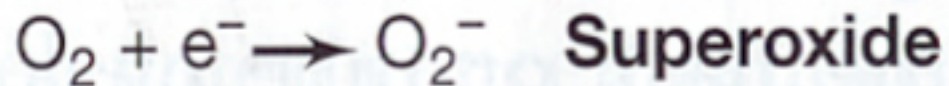
+ SOD  
- Catalase

- SOD  
- Catalase

+ SOD  
+/- Catalase  
(low levels)

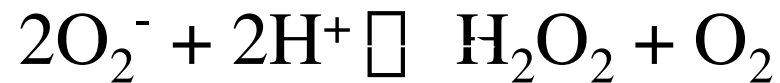
Fig. 6.15

# Toxic Forms of Oxygen



# Toxic O<sub>2</sub>-neutralizing enzymes

Superoxide Dismutase:



Catalase:

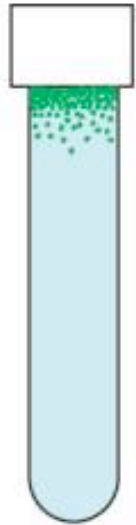


Peroxidase:



# Oxygen

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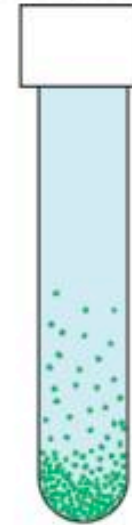
Obligate  
aerobe



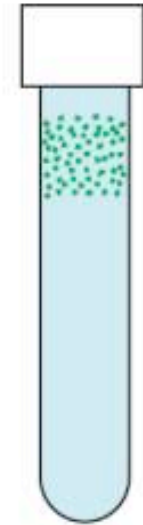
Facultative  
anaerobe



Aerotolerant  
anaerobe



Strict  
anaerobe



Microaerophile

## Enzyme content

+ SOD  
+ Catalase

+ SOD  
+ Catalase

+ SOD  
- Catalase

- SOD  
- Catalase

+ SOD  
+/- Catalase  
(low levels)

Fig. 6.15

# Anaerobic Chamber



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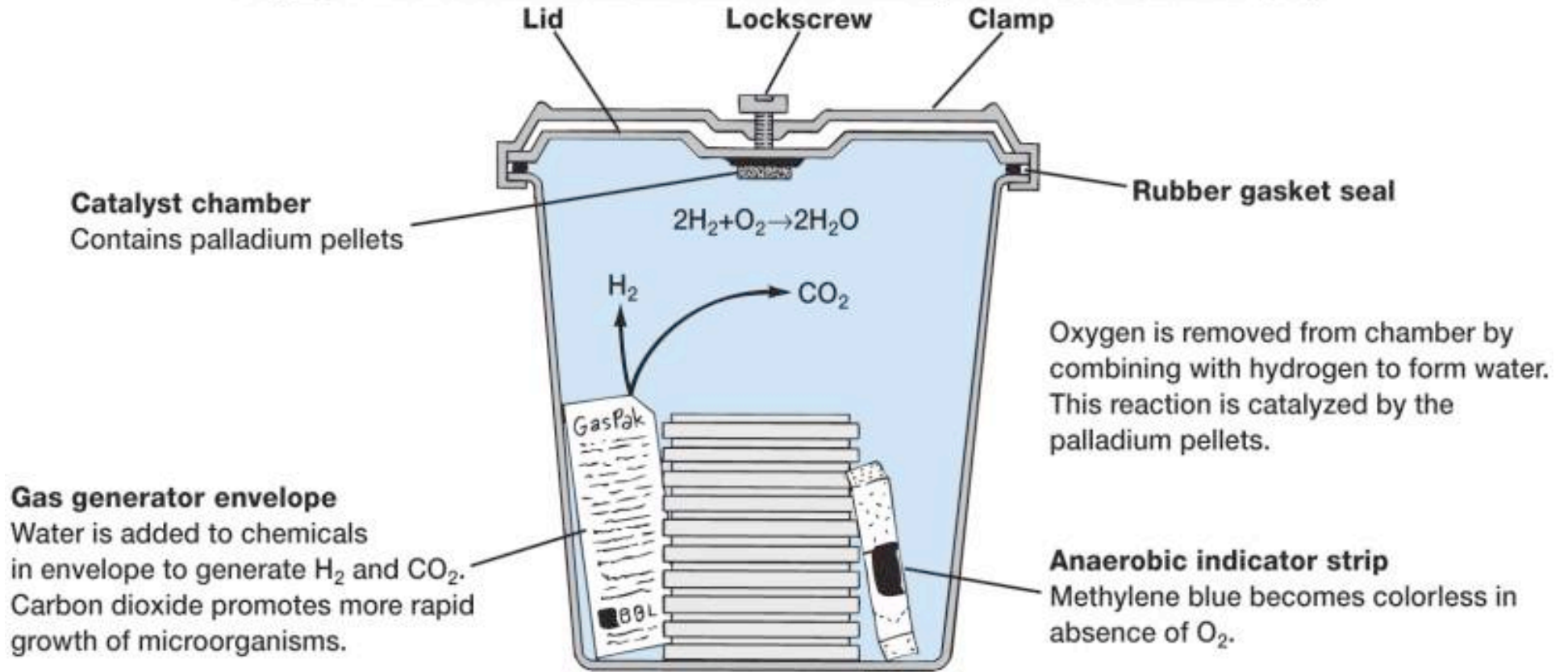


Fig. 6.17



# pH

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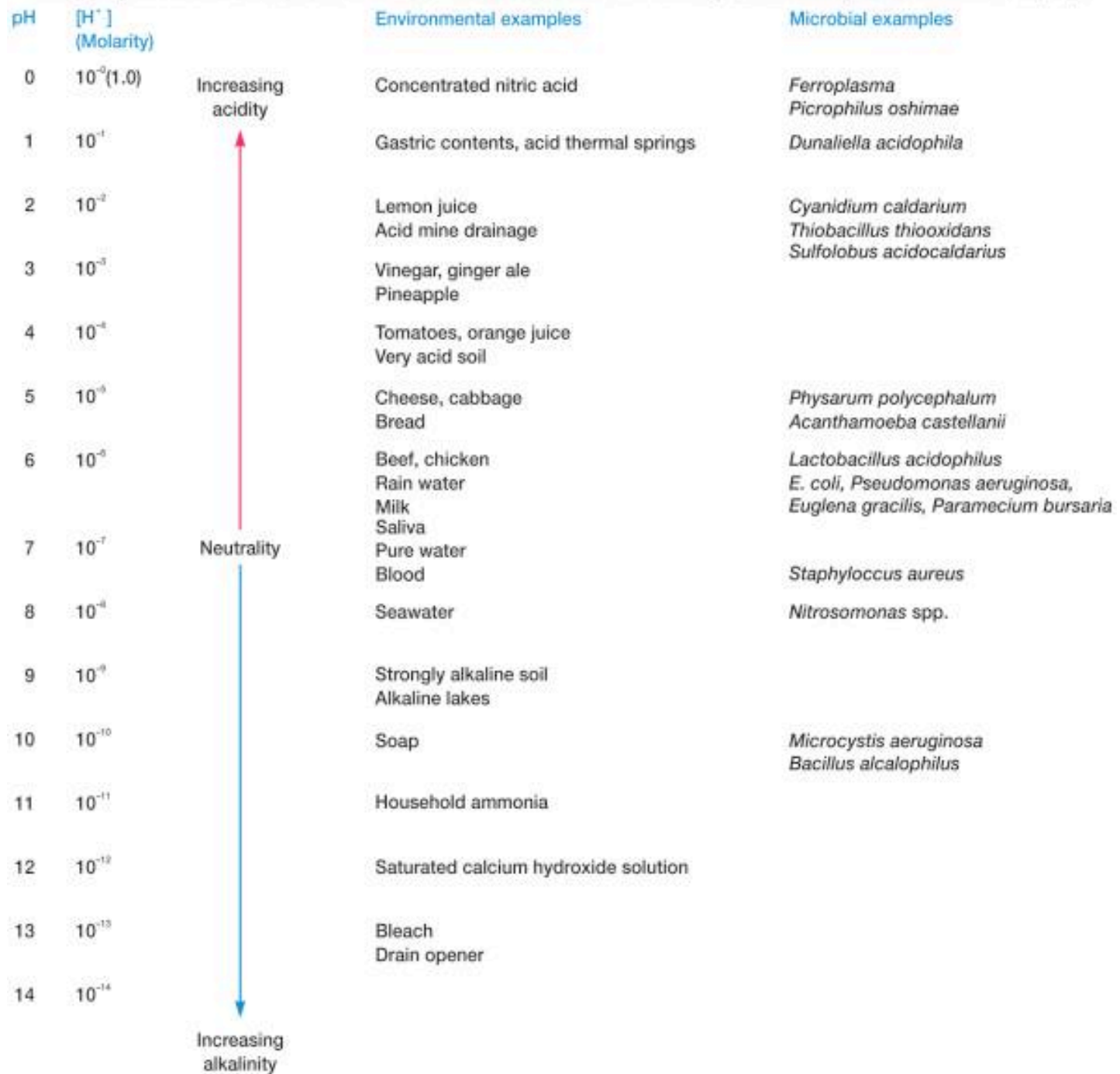


Fig. 6.12

# Osmotic pressure

- **isotonic=**
  - concentrations of solutes same on both sides membrane
- **hypotonic**
  - more water outside of cell
  - concentration of solutes greater inside cell
  - low osmotic pressure
- **hypertonic-**
  - more water inside cell
  - concentration of solutes greater outside cell
  - high osmotic pressure

# Osmotolerance

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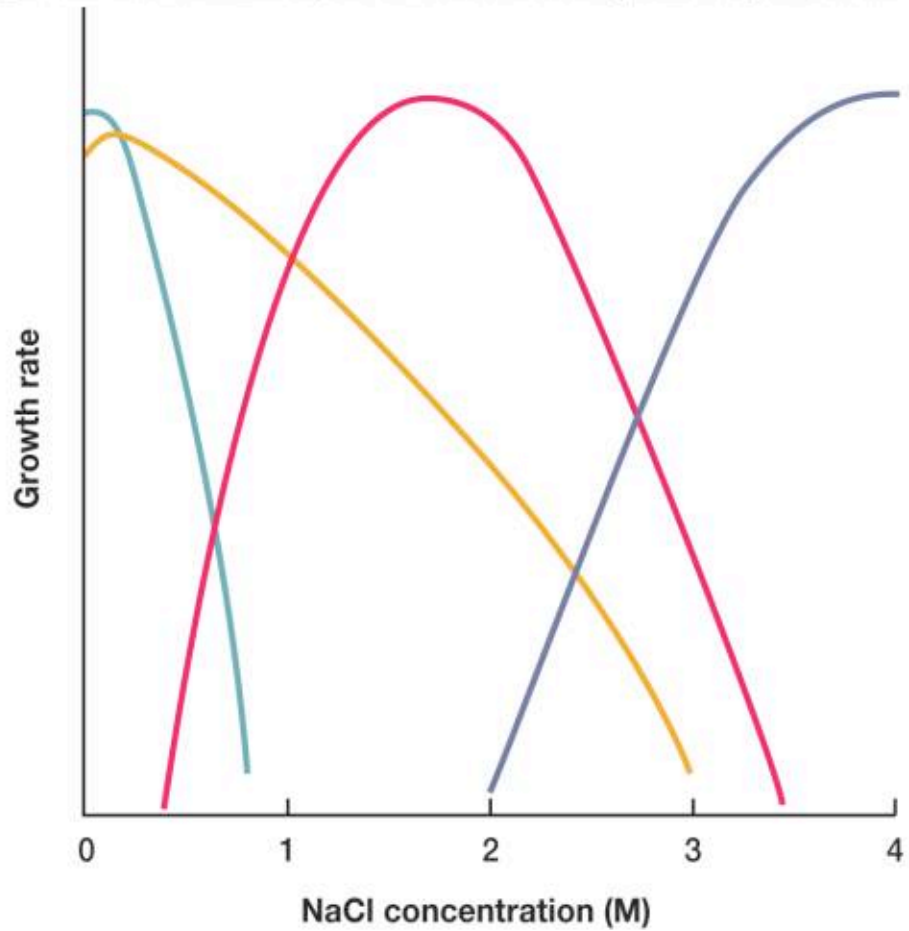


Fig. 6.11



# Osmolarity (Solute concentration)

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**Table 6.4**

**Approximate Lower  $a_w$  Limits for Microbial Growth**

Water Activity	Environment	Bacteria	Fungi	Algae
1.00—Pure water	Blood Plant wilt	Most gram-negative nonhalophiles		
0.95	Seawater			
0.90	Bread Ham	Most gram-positive rods Most cocci, <i>Bacillus</i>	<i>Basidiomycetes</i> <i>Fusarium</i> <i>Mucor</i> , <i>Rhizopus</i> Ascomycetous yeasts	Most algae
0.85	Salami	<i>Staphylococcus</i>	<i>Saccharomyces rouxii</i> (in salt)	
0.80	Preserves		<i>Penicillium</i>	
0.75	Salt lakes Salted fish	<i>Halobacterium</i> <i>Actinospora</i>	<i>Aspergillus</i>	<i>Dunaliella</i>
0.70			<i>Aspergillus</i>	
0.60	Cereals, candy, dried fruit		<i>Saccharomyces rouxii</i> (in sugars)	
	Chocolate Honey Dried milk		<i>Xeromyces bisporus</i>	
0.55—DNA disordered				

Adapted from A. D. Brown, "Microbial Water Stress," in *Bacteriological Reviews*, 40(4):803–846 1976. Copyright © 1976 by the American Society for Microbiology. Reprinted by permission.

# Chemical requirements

Carbon

Nitrogen

Sulfur

Phosphorous

Potassium

Magnesium

Calcium

Trace elements:

iron, copper, zinc

**Table 5.4**  
**Examples of Defined Media**

<b>BG-11 Medium for Cyanobacteria</b>	<b>Amount (g/liter)</b>
NaNO <sub>3</sub>	1.5
K <sub>2</sub> HPO <sub>4</sub> · 3H <sub>2</sub> O	0.04
MgSO <sub>4</sub> · 7H <sub>2</sub> O	0.075
CaCl <sub>2</sub> · 2H <sub>2</sub> O	0.036
Citric acid	0.006
Ferric ammonium citrate	0.006
EDTA (Na <sub>2</sub> Mg salt)	0.001
Na <sub>2</sub> CO <sub>3</sub>	0.02
Trace metal solution <sup>a</sup>	1.0 ml/liter
Final pH 7.4	
<b>Medium for <i>Escherichia coli</i></b>	<b>Amount (g/liter)</b>
Glucose	1.0
Na <sub>2</sub> HPO <sub>4</sub>	16.4
KH <sub>2</sub> PO <sub>4</sub>	1.5
(NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub>	2.0
MgSO <sub>4</sub> · 7H <sub>2</sub> O	200.0 mg
CaCl <sub>2</sub>	10.0 mg
FeSO <sub>4</sub> · 7H <sub>2</sub> O	0.5 mg
Final pH 6.8–7.0	

**Sources:** Data from Rippka, et al. *Journal of General Microbiology*, 111:1–61, 1979; and S. S. Cohen, and R. Arbogast, *Journal of Experimental Medicine*, 91:619, 1950.

<sup>a</sup>The trace metal solution contains H<sub>3</sub>BO<sub>3</sub>, MnCl<sub>2</sub> · 4H<sub>2</sub>O, ZnSO<sub>4</sub> · 7H<sub>2</sub>O, Na<sub>2</sub>Mo<sub>4</sub> · 2H<sub>2</sub>O, CuSO<sub>4</sub> · 5H<sub>2</sub>O, and Co(NO<sub>3</sub>)<sub>2</sub> · 6H<sub>2</sub>O.

**Table 5.5****Some Common Complex Media**

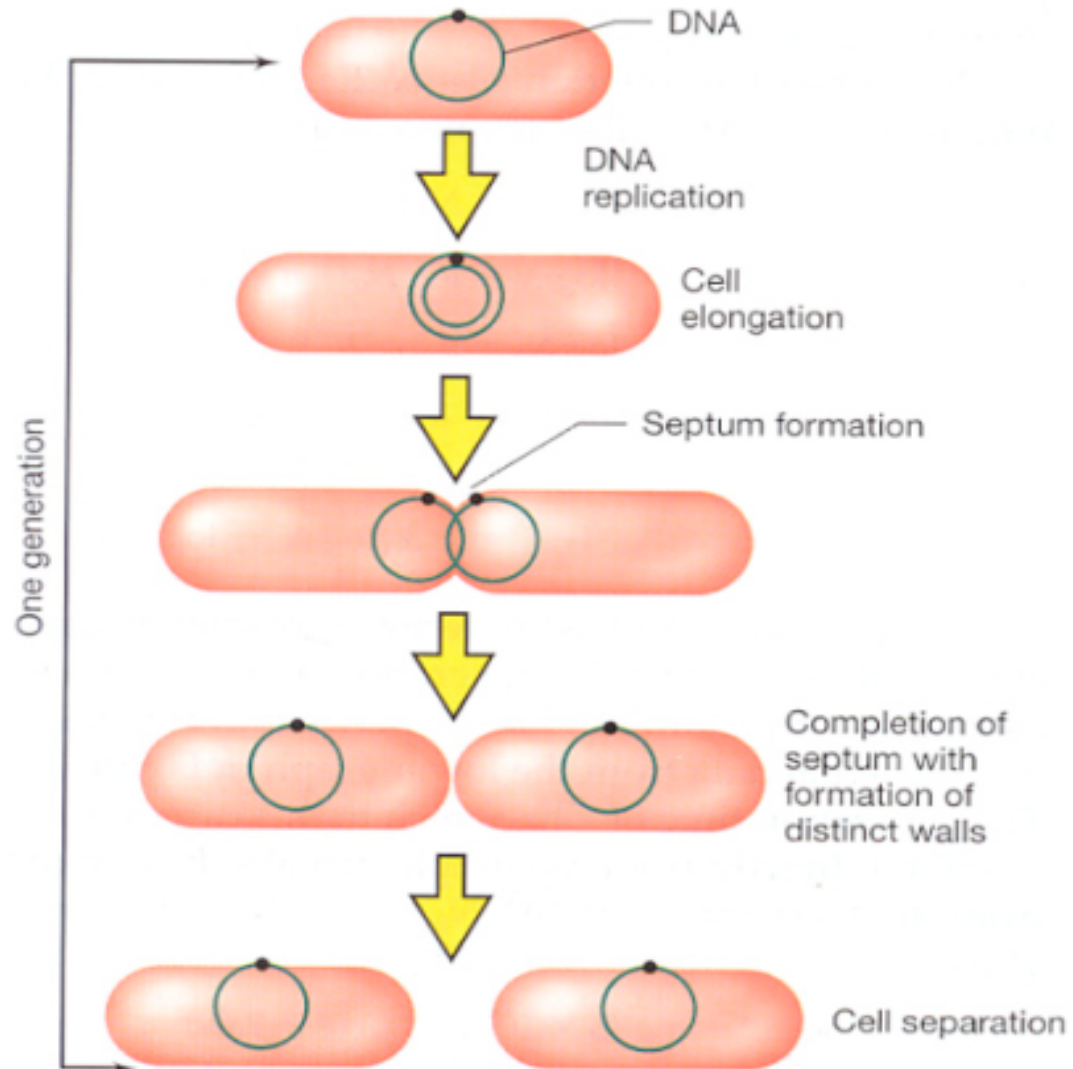
<b>Nutrient Broth</b>	<b>Amount (g/liter)</b>
Peptone (gelatin hydrolysate)	5
Beef extract	3
<b>Tryptic Soy Broth</b>	
Tryptone (pancreatic digest of casein)	17
Peptone (soybean digest)	3
Glucose	2.5
Sodium chloride	5
Dipotassium phosphate	2.5
<b>MacConkey Agar</b>	
Pancreatic digest of gelatin	17.0
Pancreatic digest of casein	1.5
Peptic digest of animal tissue	1.5
Lactose	10.0
Bile salts	1.5
Sodium chloride	5.0
Neutral red	0.03
Crystal violet	0.001
Agar	13.5

# Other types of media- check them out in lab!

- Selective
  - suppresses growth of unwanted bacteria
  - encourages growth of desired bacteria
- Differential
  - distinguish colonies of desired organism from other colonies
- Enrichment
  - increase number of organisms
    - (few-> many)



# Binary fission



# Microbial Growth Curve

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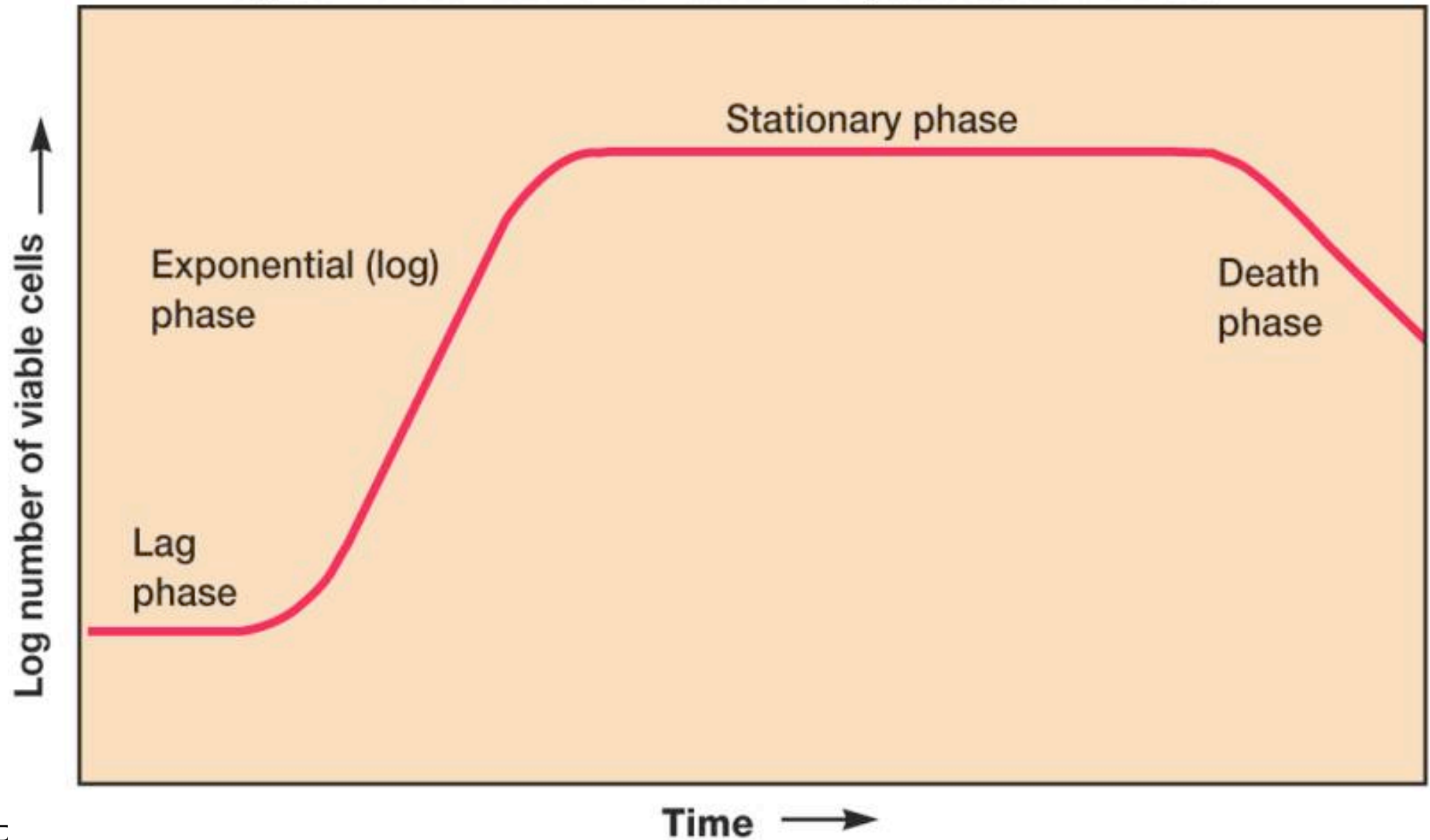


Fig. 0.1

# Exponential Growth

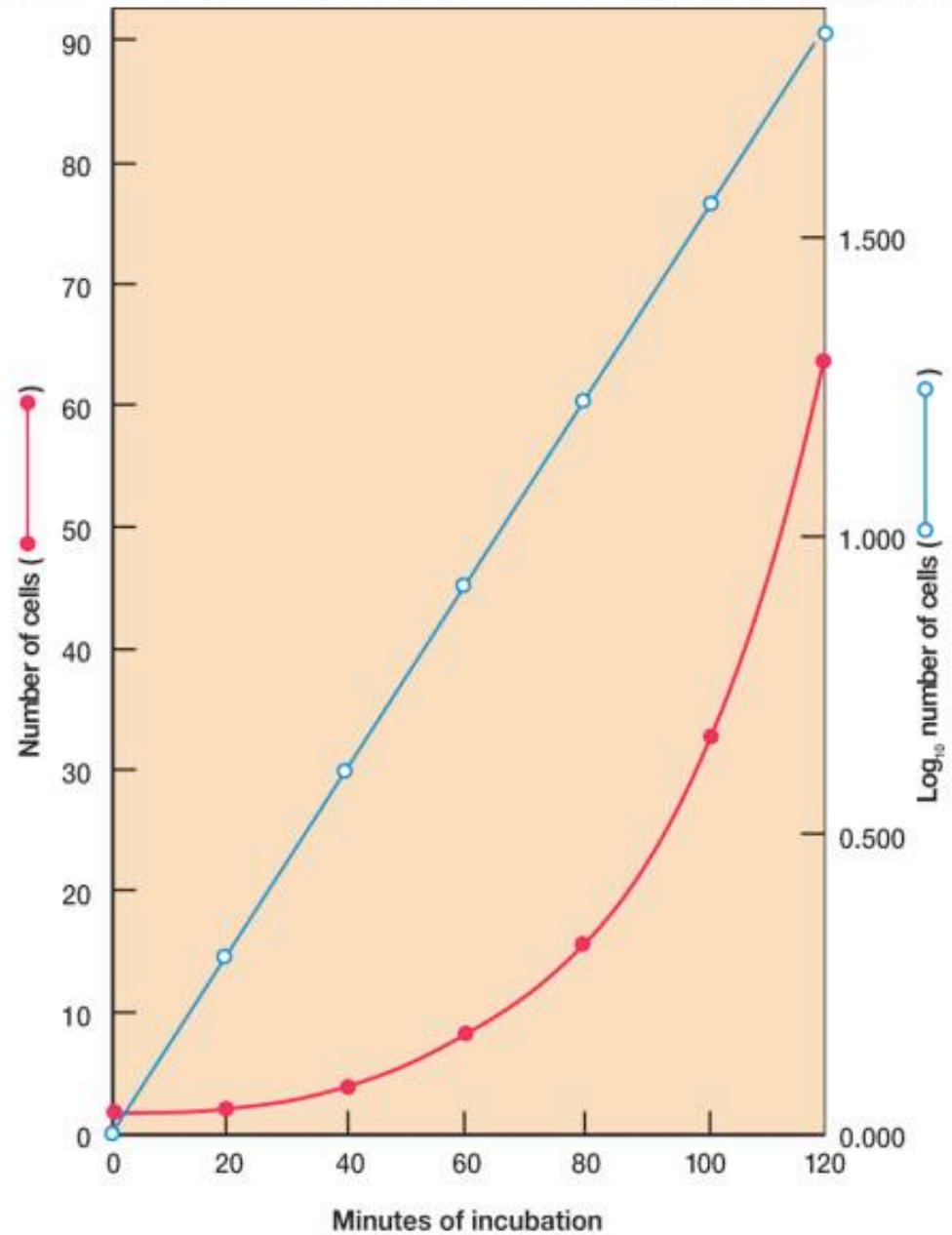


Fig. 6.3

**Table 6.1**

**An Example of Exponential Growth**

<b>Time<sup>a</sup></b>	<b>Division Number</b>	<b><math>2^n</math></b>	<b>Population (<math>N_o \times 2^n</math>)</b>	<b><math>\log_{10}N_t</math></b>
0	0	$2^0 = 1$	1	0.000
20	1	$2^1 = 2$	2	0.301
40	2	$2^2 = 4$	4	0.602
60	3	$2^3 = 8$	8	0.903
80	4	$2^4 = 16$	16	1.204
100	5	$2^5 = 32$	32	1.505
120	6	$2^6 = 64$	64	1.806

<sup>a</sup>The hypothetical culture begins with one cell having a 20-minute generation time.

# Generation time determination

$$N_t = N_o \times 2^n$$

$$n = \log N_t - \log N_o / \log 2$$

$$k = n/t$$

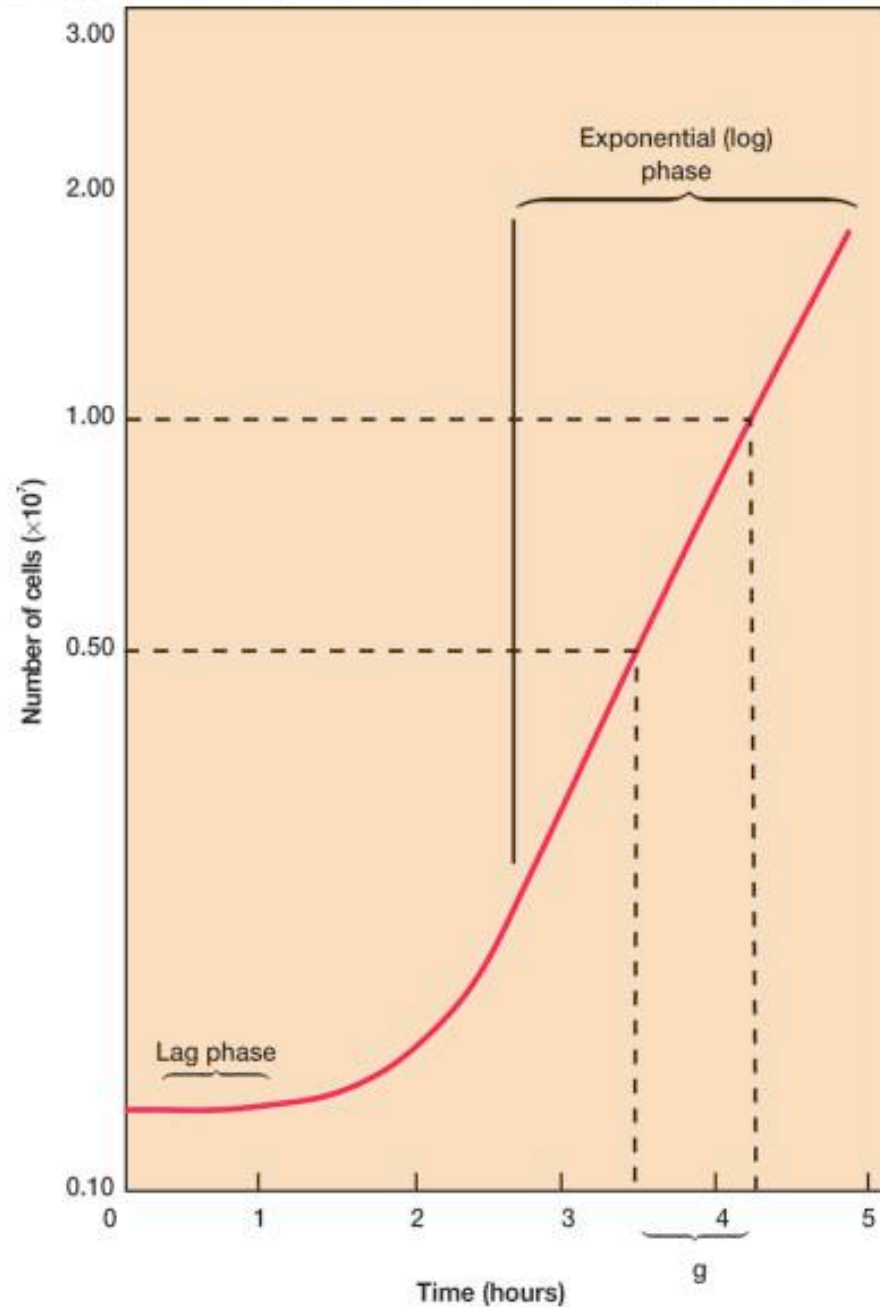


Fig. 6.4

**Table 6.2**  
**Generation Times for Selected Microorganisms**

Microorganism	Temperature (°C)	Generation Time (Hours)
<b>Bacteria</b>		
<i>Beneckea natriegens</i>	37	0.16
<i>Escherichia coli</i>	40	0.35
<i>Bacillus subtilis</i>	40	0.43
<i>Staphylococcus aureus</i>	37	0.47
<i>Pseudomonas aeruginosa</i>	37	0.58
<i>Clostridium botulinum</i>	37	0.58
<i>Rhodospirillum rubrum</i>	25	4.6–5.3
<i>Anabaena cylindrica</i>	25	10.6
<i>Mycobacterium tuberculosis</i>	37	≈12
<i>Treponema pallidum</i>	37	33
<b>Algae</b>		
<i>Scenedesmus quadricauda</i>	25	5.9
<i>Chlorella pyrenoidosa</i>	25	7.75
<i>Asterionella formosa</i>	20	9.6
<i>Euglena gracilis</i>	25	10.9
<i>Ceratium tripos</i>	20	82.8
<b>Protozoa</b>		
<i>Tetrahymena geleii</i>	24	2.2–4.2
<i>Leishmania donovani</i>	26	10–12
<i>Paramecium caudatum</i>	26	10.4
<i>Acanthamoeba castellanii</i>	30	11–12
<i>Giardia lamblia</i>	37	18
<b>Fungi</b>		
<i>Saccharomyces cerevisiae</i>	30	2
<i>Monilinia fructicola</i>	25	30

# Chemostat Continuous Culture

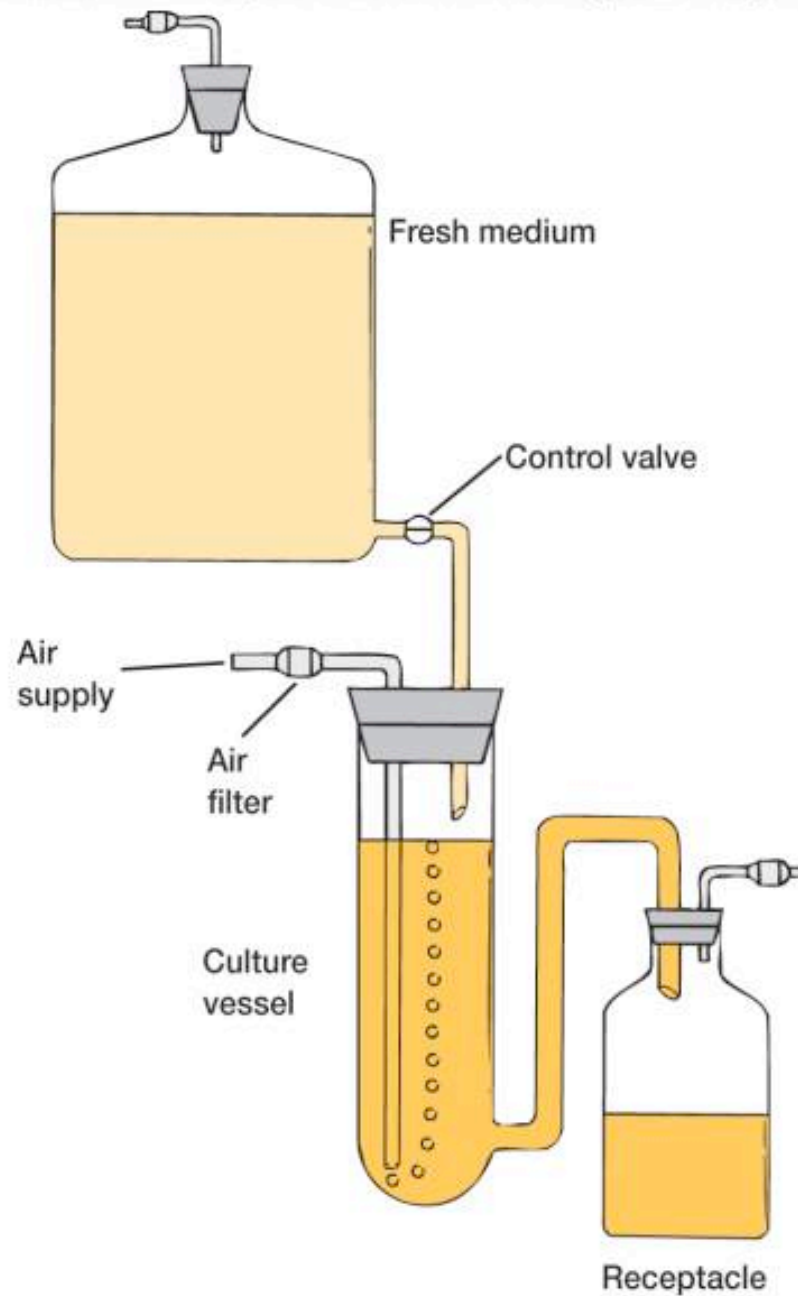


Fig. 6.9

# Turbidity(Absorbance)

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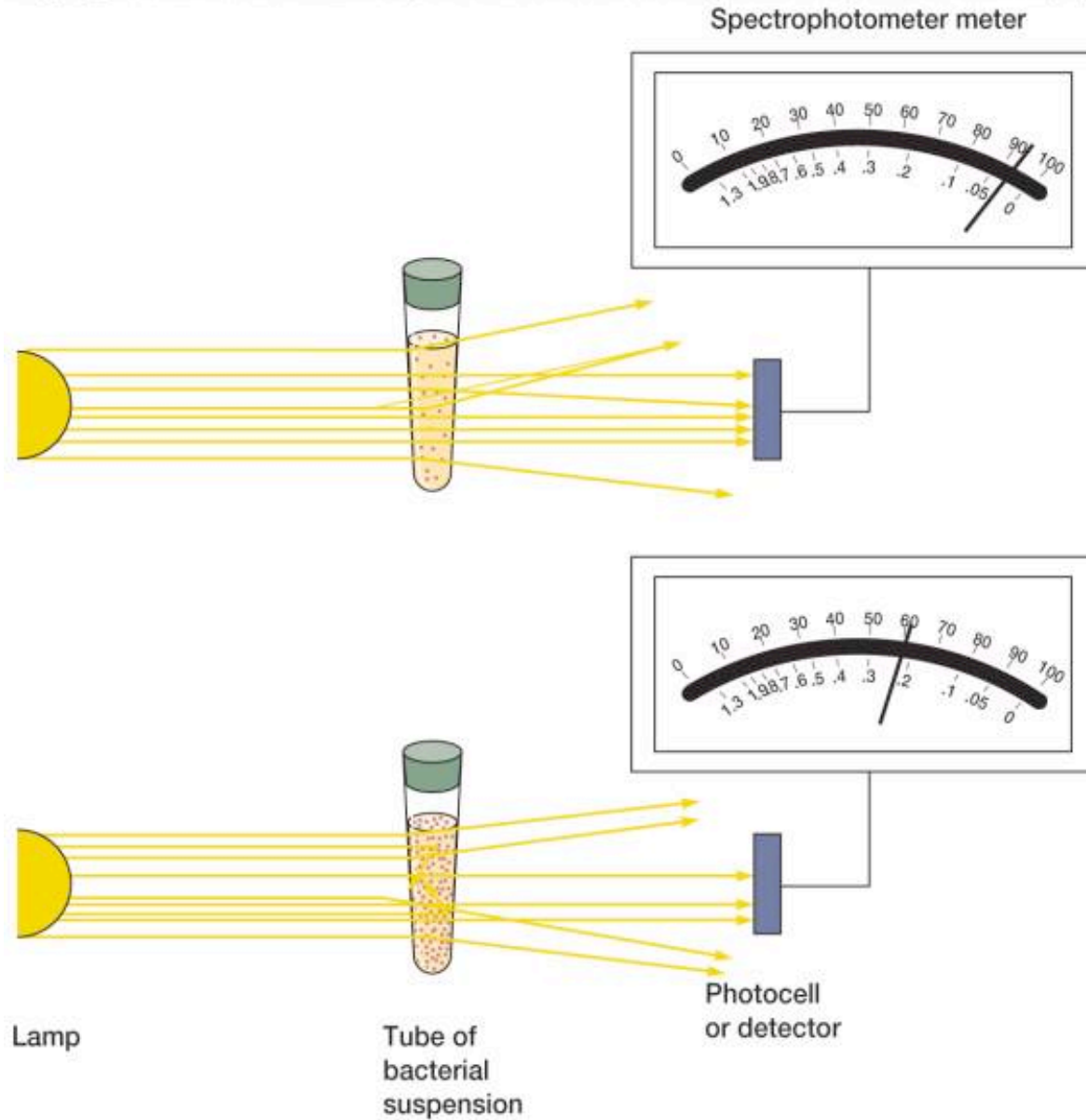
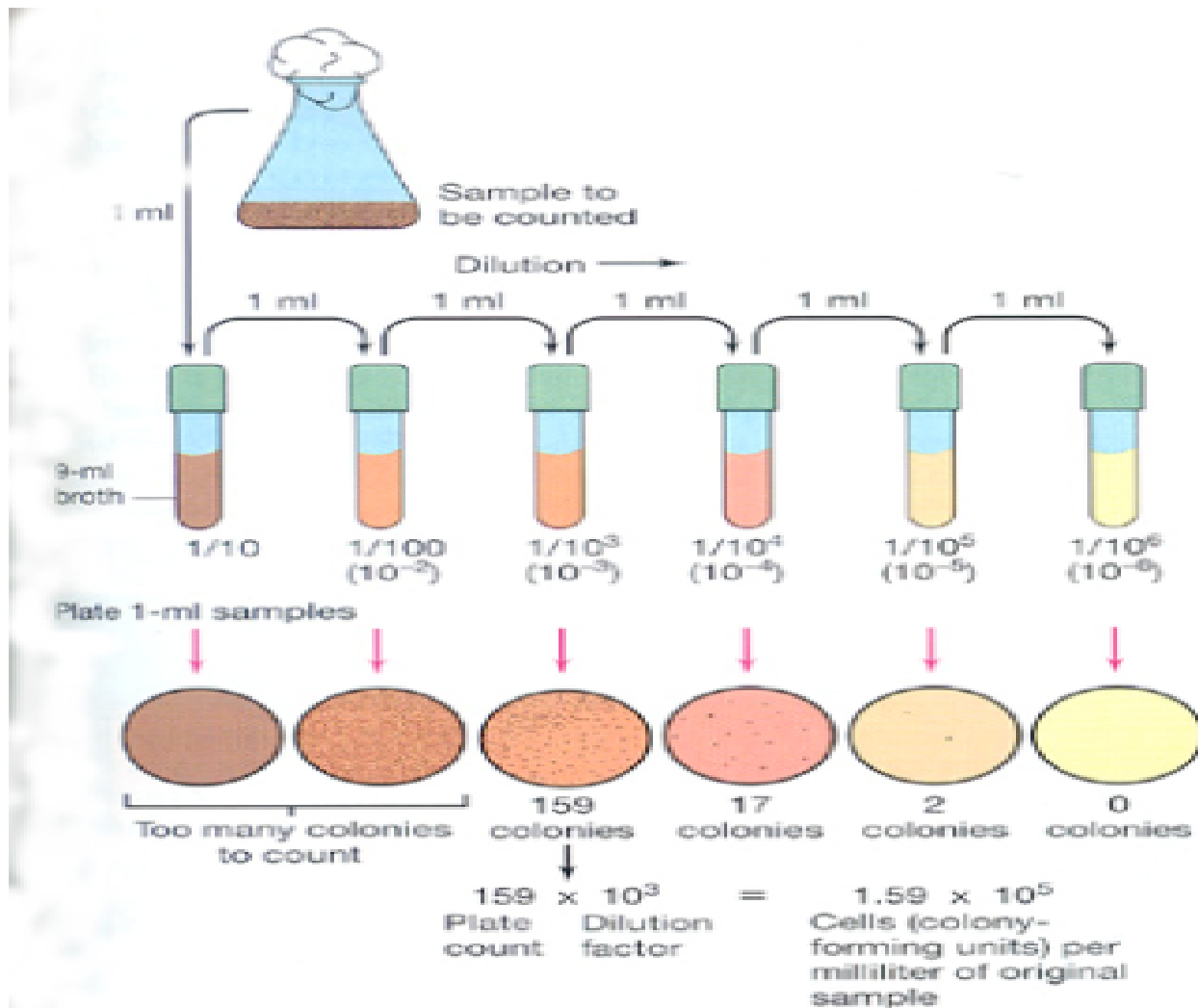


Fig. 6.8

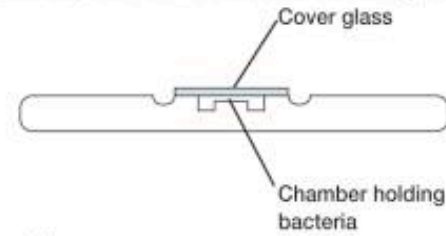


# Viabie cell count

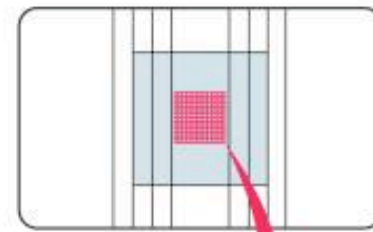


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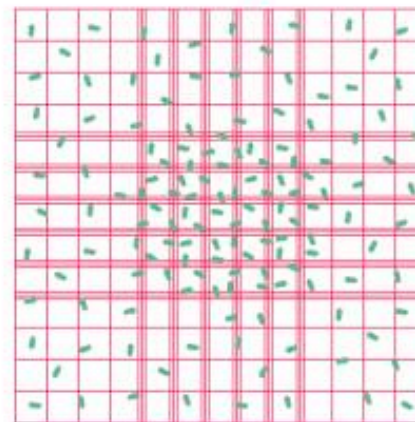
# Direct cell count



(a)



(b)



(c)

Fig. 6.5

# Membrane filtration

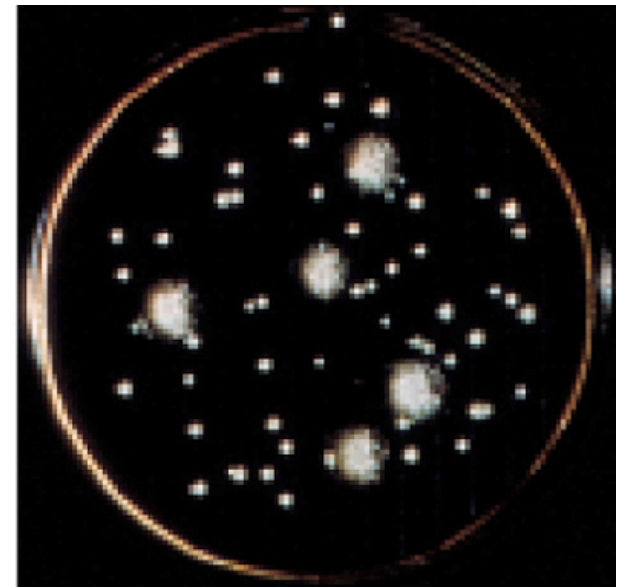
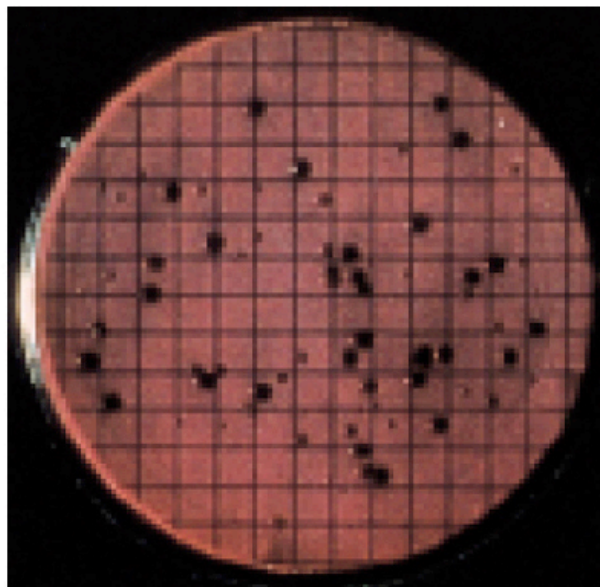
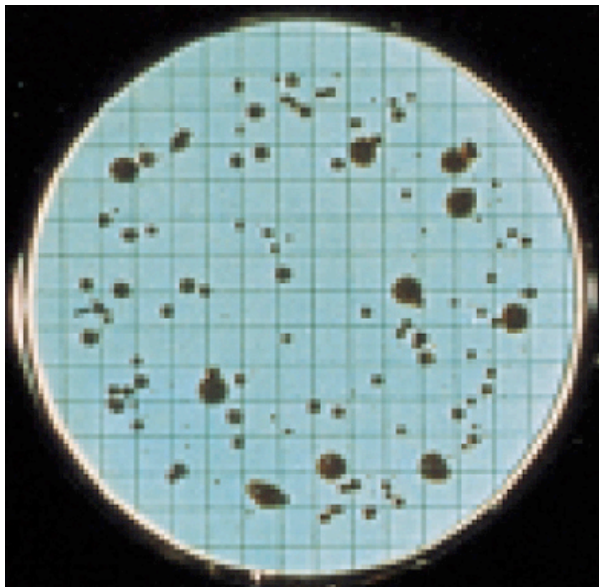
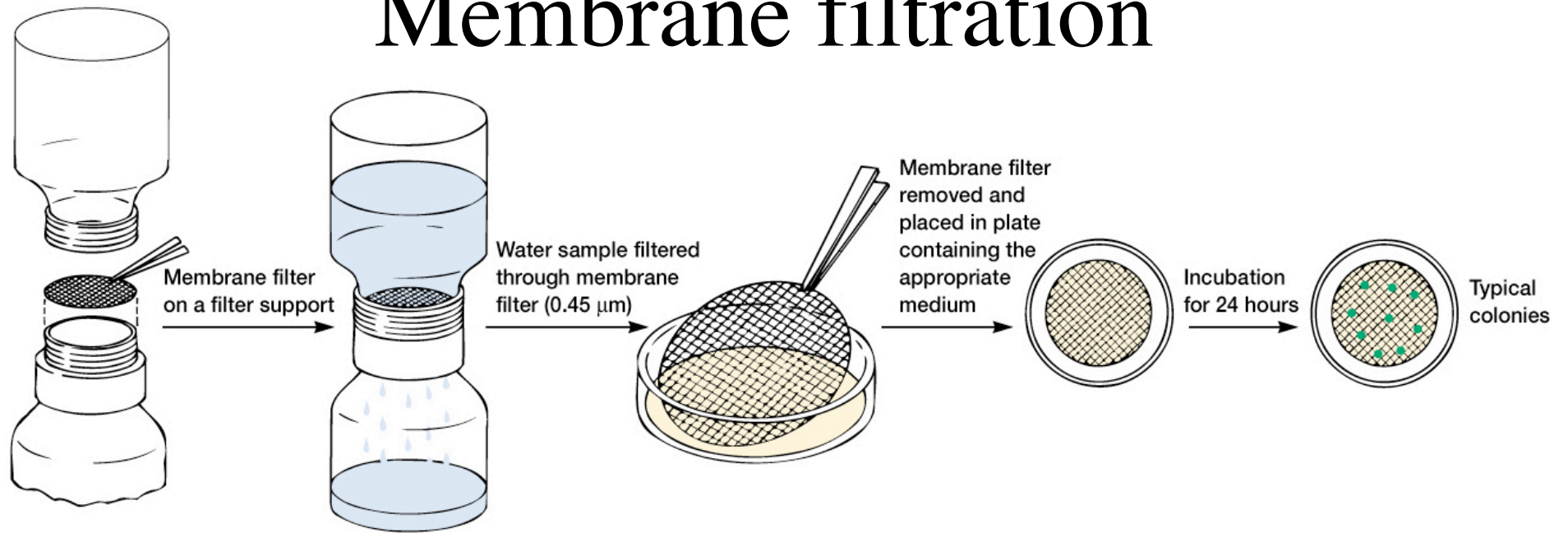


Fig. 6.6