

Morphological and physiological characteristics used in classification

- Colony morphology
- Colony color
- Cell shape
- Staining results (Gram)
- Endospore
- Motility
- Flagella
- Size
- Cellular inclusions
- Ultrastructure
- Oxygen relationship
- Energy sources
- Electron sources
- Carbon sources
- Fermentation products
- Cell wall components
- Temp and pH optima
- Osmotic tolerance
- Photosynthetic mechanism
- Secondary metabolites
- Sensitivity to antibiotics

Oxygen requirements

- Aerobes
- Anaerobes
- Microaerophiles
- Obligate aerobes
- Obligate anaerobes
- Facultative anaerobes

Table 5.1

Sources of Carbon, Energy, and Electrons

Carbon Sources

Autotrophs	CO ₂ sole or principal biosynthetic carbon source (<i>pp. 202–3</i>) ^a
Heterotrophs	Reduced, preformed, organic molecules from other organisms (<i>chapters 9 and 10</i>)

Energy Sources

Phototrophs	Light (<i>pp. 190–96</i>)
Chemotrophs	Oxidation of organic or inorganic compounds (<i>chapter 9</i>)

Electron Sources

Lithotrophs	Reduced inorganic molecules (<i>pp. 188–90</i>)
Organotrophs	Organic molecules (<i>chapter 9</i>)

^aFor each category, the location of material describing the participating metabolic pathways is given within the parentheses.

Table 5.2
Major Nutritional Types of Microorganisms

Major Nutritional Types ^a	Sources of Energy, Hydrogen/Electrons, and Carbon	Representative Microorganisms
Photolithotrophic autotrophy (Photolithoautotrophy)	Light energy Inorganic hydrogen/electron (H/e^-) donor CO_2 carbon source	Algae Purple and green sulfur bacteria Cyanobacteria
Photoorganotrophic heterotrophy (Photoorganoheterotrophy)	Light energy Organic H/e^- donor Organic carbon source (CO_2 may also be used)	Purple nonsulfur bacteria Green nonsulfur bacteria
Chemolithotrophic autotrophy (Chemolithoautotrophy)	Chemical energy source (inorganic) Inorganic H/e^- donor CO_2 carbon source	Sulfur-oxidizing bacteria Hydrogen bacteria Nitrifying bacteria Iron-oxidizing bacteria
Chemoorganotrophic heterotrophy (Chemoorganoheterotrophy)	Chemical energy source (organic) Organic H/e^- donor Organic carbon source	Protozoa Fungi Most nonphotosynthetic bacteria (including most pathogens)

^aBacteria in other nutritional categories have been found. The categories are defined in terms of energy, electron, and carbon sources. Condensed versions of these names are given in parentheses.

Numerical Taxonomy

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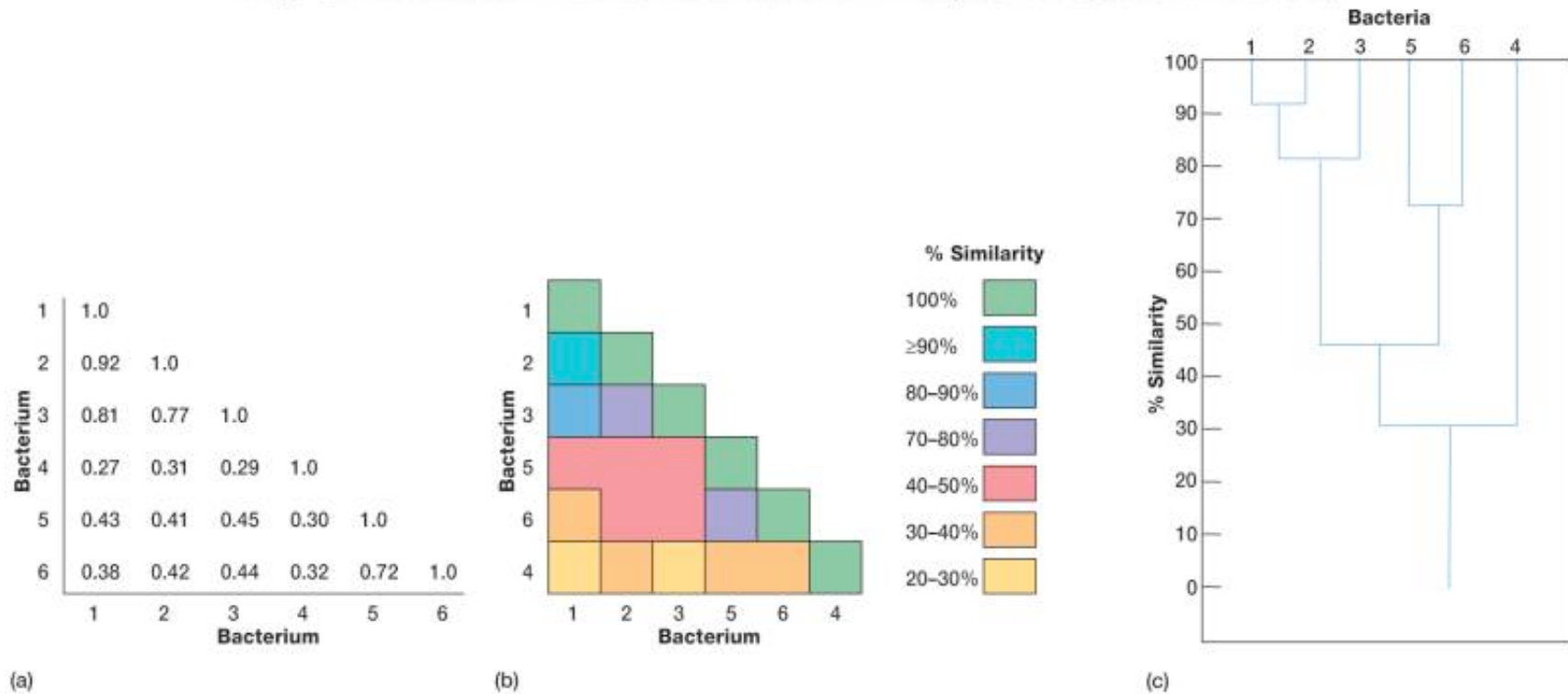


Fig. 19.5

Molecular characteristics used in classification

- Nucleic acid base composition (G+C %)
- Nucleic acid hybridization
- Nucleic acid sequencing
 - 16S rRNA
 - Signature sequences
- Nucleic acid/protein sequence comparisons
- Complete genome sequencing

DNA melting curve (G+C %)

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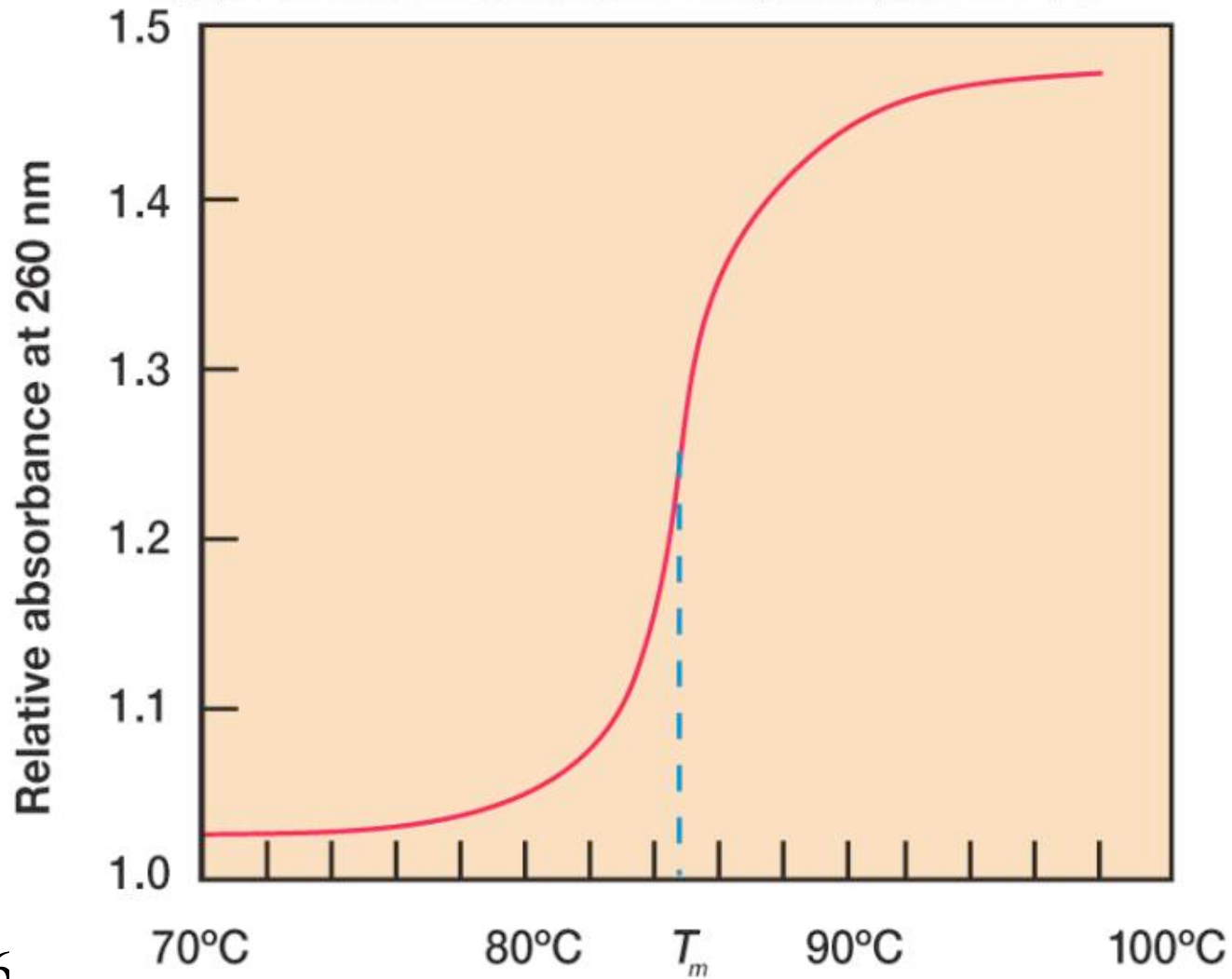


Fig. 19.6

Table 19.5**Representative G + C Contents of Microorganisms**

Organism	Percent G + C	Organism	Percent G + C	Organism	Percent G + C
Bacteria		<i>Salmonella</i>	50–53	<i>Plasmodium berghei</i>	41
<i>Actinomyces</i>	59–73	<i>Spirillum</i>	38	<i>Stentor polymorphus</i>	45
<i>Anabaena</i>	38–44	<i>Spirochaeta</i>	51–65	<i>Tetrahymena</i>	19–33
<i>Bacillus</i>	32–62	<i>Staphylococcus</i>	30–38	<i>Trichomonas</i>	29–34
<i>Bacteroides</i>	28–61	<i>Streptococcus</i>	33–44	<i>Trypanosoma</i>	45–59
<i>Bdellovibrio</i>	33–52	<i>Streptomyces</i>	69–73		
<i>Caulobacter</i>	63–67	<i>Sulfolobus</i>	31–37	Slime Molds	
<i>Chlamydia</i>	41–44	<i>Thermoplasma</i>	46	<i>Dictyostelium</i>	22–25
<i>Chlorobium</i>	49–58	<i>Thiobacillus</i>	52–68	<i>Lycogala</i>	42
<i>Chromatium</i>	48–70	<i>Treponema</i>	25–54	<i>Physarum polycephalum</i>	38–42
<i>Clostridium</i>	21–54				
<i>Cytophaga</i>	33–42	Algae		Fungi	
<i>Deinococcus</i>	62–70	<i>Acetabularia mediterranea</i>	37–53	<i>Agaricus bisporus</i>	44
<i>Escherichia</i>	48–52	<i>Chlamydomonas</i>	60–68	<i>Amanita muscaria</i>	57
<i>Halobacterium</i>	66–68	<i>Chlorella</i>	43–79	<i>Aspergillus niger</i>	52
<i>Hyphomicrobium</i>	59–67	<i>Cyclotella cryptica</i>	41	<i>Blastocladiella emersonii</i>	66
<i>Methanobacterium</i>	32–50	<i>Euglena gracilis</i>	46–55	<i>Candida albicans</i>	33–35
<i>Micrococcus</i>	64–75	<i>Nitella</i>	49	<i>Claviceps purpurea</i>	53
<i>Mycobacterium</i>	62–70	<i>Nitzschia angularis</i>	47	<i>Coprinus lagopus</i>	52–53
<i>Mycoplasma</i>	23–40	<i>Ochromonas danica</i>	48	<i>Fomes fraxineus</i>	56
<i>Myxococcus</i>	68–71	<i>Peridinium triquetrum</i>	53	<i>Mucor rouxii</i>	38
<i>Neisseria</i>	47–54	<i>Scenedesmus</i>	52–64	<i>Neurospora crassa</i>	52–54
<i>Nitrobacter</i>	60–62	<i>Spirogyra</i>	39	<i>Penicillium notatum</i>	52
<i>Oscillatoria</i>	40–50	<i>Volvox carteri</i>	50	<i>Polyporus palustris</i>	56
<i>Prochloron</i>	41			<i>Rhizopus nigricans</i>	47
<i>Proteus</i>	38–41	Protozoa		<i>Saccharomyces cerevisiae</i>	36–42
<i>Pseudomonas</i>	58–70	<i>Acanthamoeba castellanii</i>	56–58	<i>Saprolegnia parasitica</i>	61
<i>Rhodospirillum</i>	62–66	<i>Amoeba proteus</i>	66		
<i>Rickettsia</i>	29–33	<i>Paramecium</i> spp.	29–39		

Nucleic Acid Hybridization

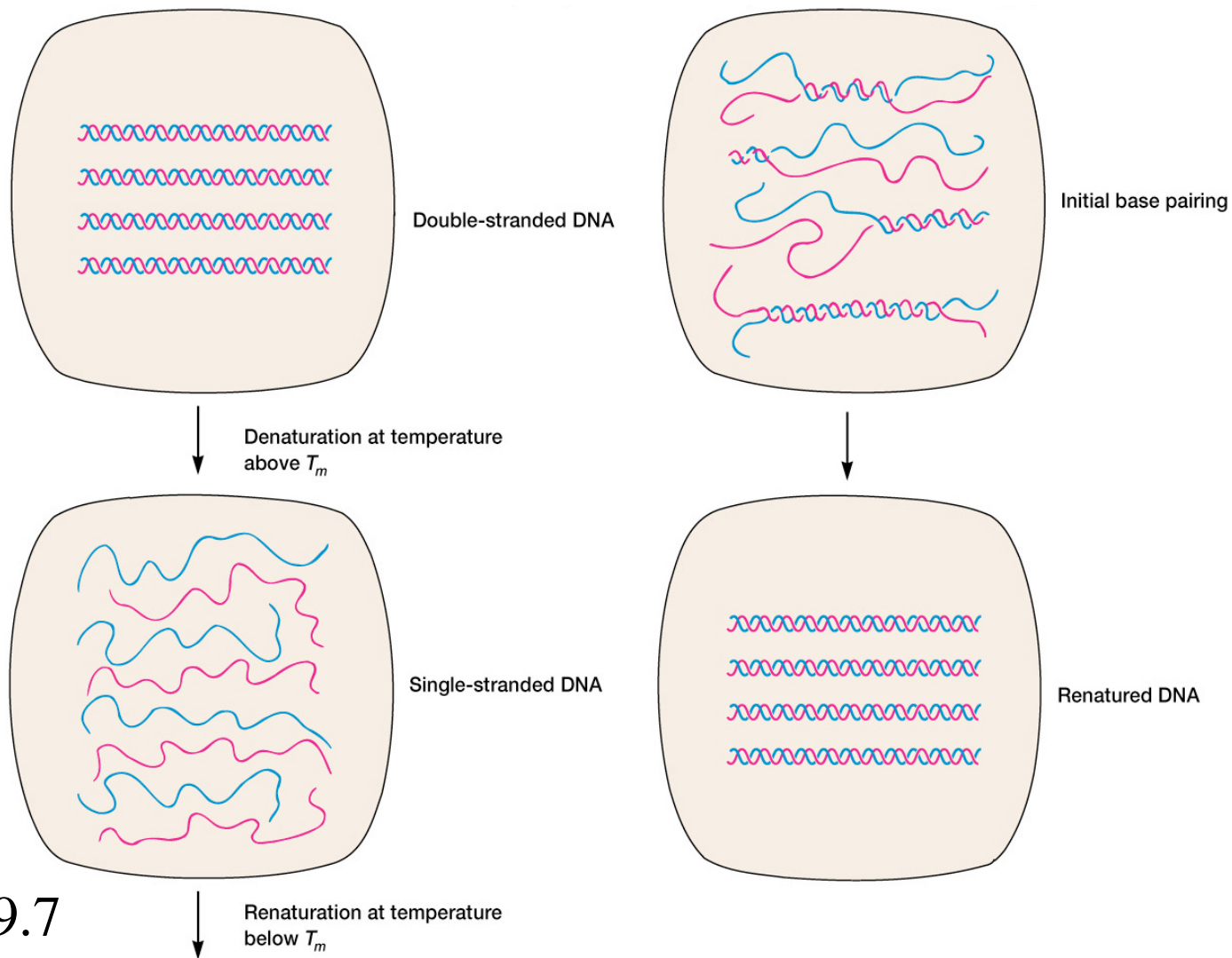


Fig. 19.7

Hybridization based on complementarity of DNA sequences

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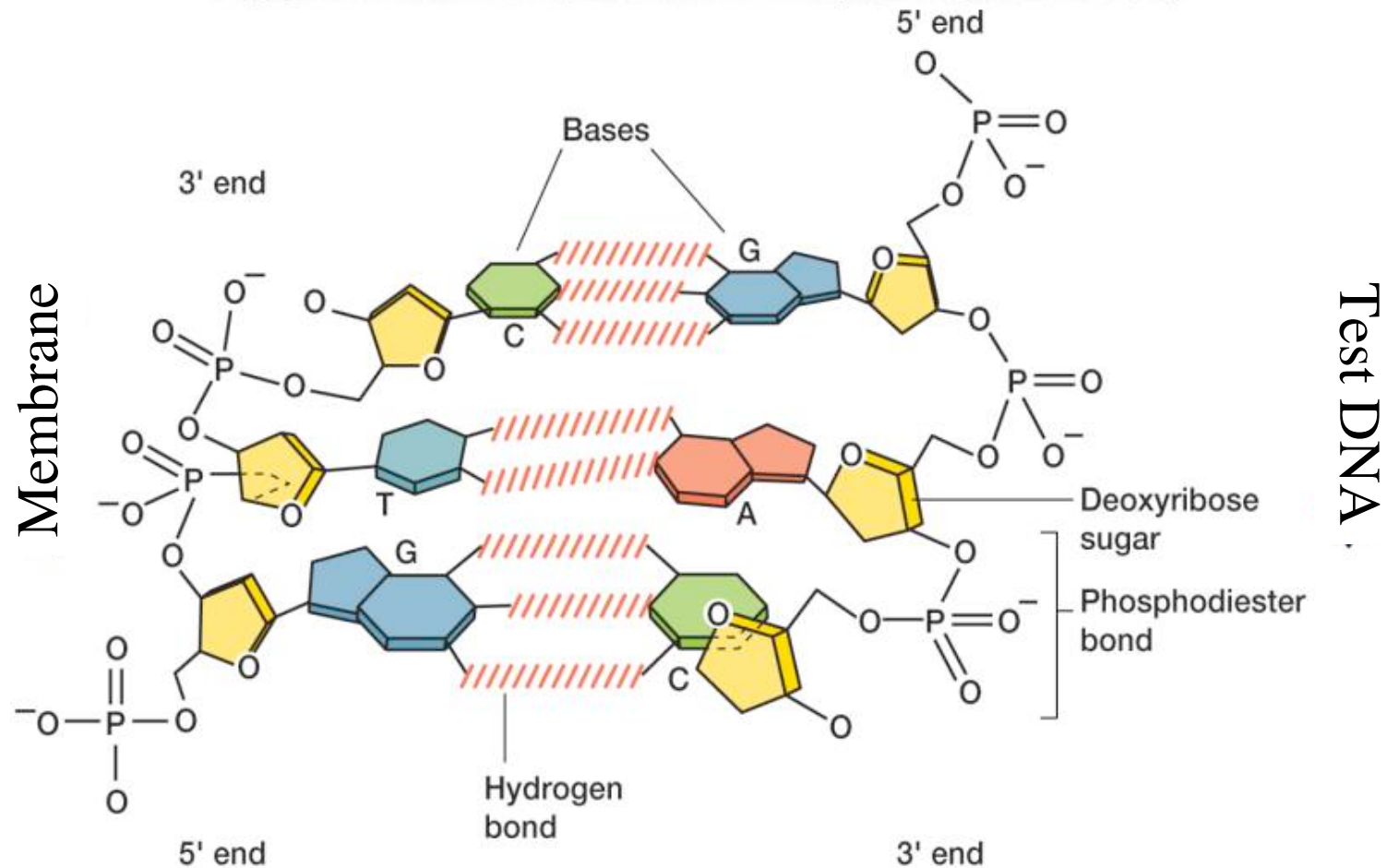


Table 19.6

Comparison of *Neisseria* Species by DNA Hybridization Experiments

Membrane-Attached DNA ^a	Percent Homology ^b
<i>Neisseria meningitidis</i>	100
<i>N. gonorrhoeae</i>	78
<i>N. sicca</i>	45
<i>N. flava</i>	35

Source: Data from T. E. Staley and R. R. Colwell, “Applications of Molecular Genetics and Numerical Taxonomy to the Classification of Bacteria” in *Annual Review of Ecology and Systematics*, 8: 282, 1973.

^aThe experimental membrane-attached nonradioactive DNA from each species was incubated with radioactive *N. meningitidis* DNA, and the amount of radioactivity bound to the membrane was measured. The more radioactivity bound, the greater the homology between DNA sequences.

^b
$$\frac{\text{N.meningitidis DNA bound to experimental DNA}}{\text{Amount bound to membrane attached N.meningitidis DNA}} \times 100$$

Small ribosomal subunit RNA

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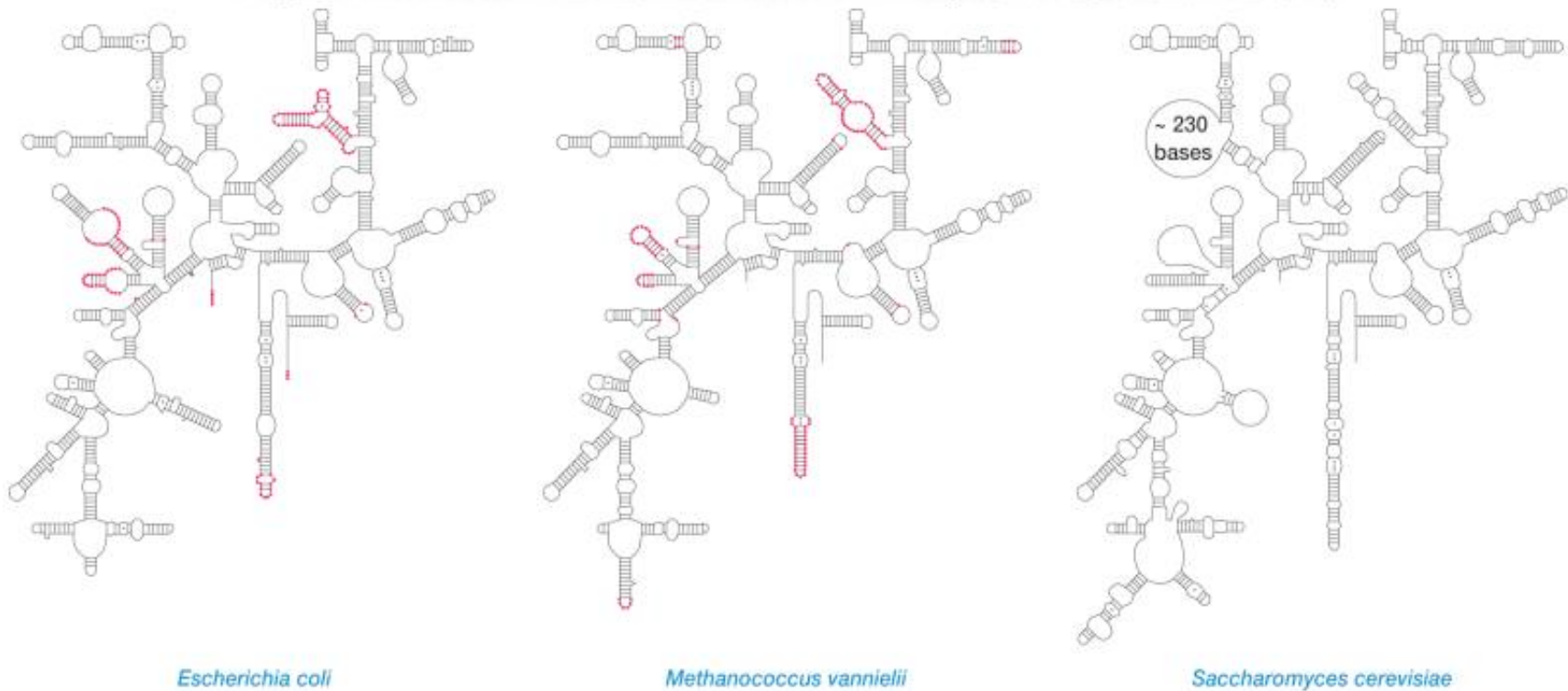


Fig. 19.9

Table 19.7**Selected 16S rRNA Signature Sequences for Some Bacterial Groups^a**

Position in rRNA	Consensus Composition	γ -Proteobacteria	Cyanobacteria	Spirochetes	<i>Bacteroides</i>	Green Sulfur	Green Nonsulfur	<i>Deinococcus</i>	Gram Positive (Low GC)	Gram Positive (High GC)	<i>Planctomyces</i>
47	C	+	+	U	+	+	+	+	+	+	G
53	A	+	+	G	+	+	G	+	+	+	G
570	G	+	+	+	U	+	+	+	+	+	U
812	G	c	+	+	+	+	+	C	+	+	+
906	G	Ag	+	+	+	+	A	+	+	A	+
955	U	+	+	+	+	+	+	+	+	AC	C
1,207	G	+	C	+	+	+	+	+	C	C	+
1,234	C	+	+	a	U	A	+	+	+	+	+

^aA plus sign in a column means that the group has the same base as the consensus sequence. If the letter is given in upper case, it is changed in more than 90% of the cases. A lowercase letter signifies a minor occurrence base (<15% of the cases).

DNA sequencing

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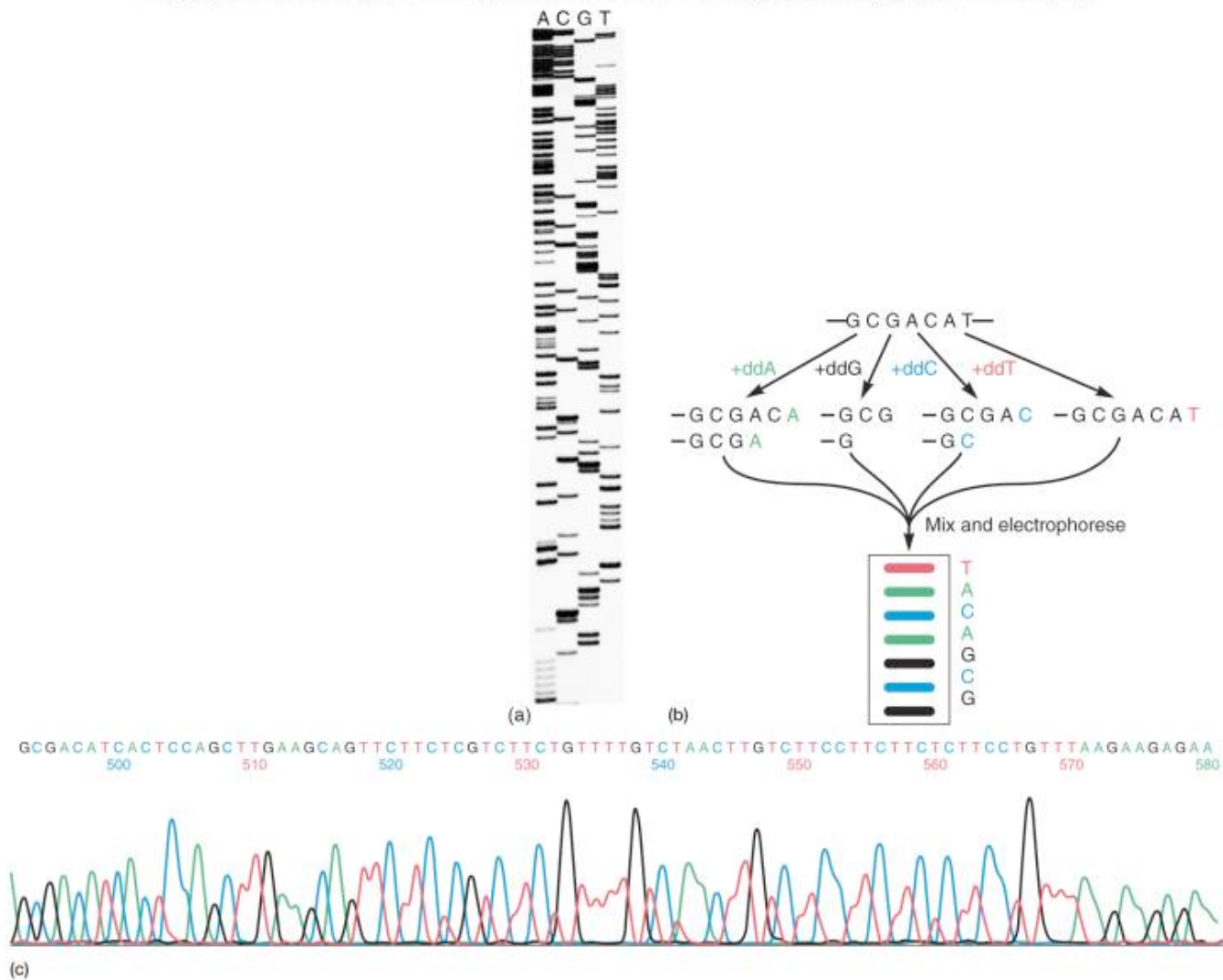


Fig. 15.2

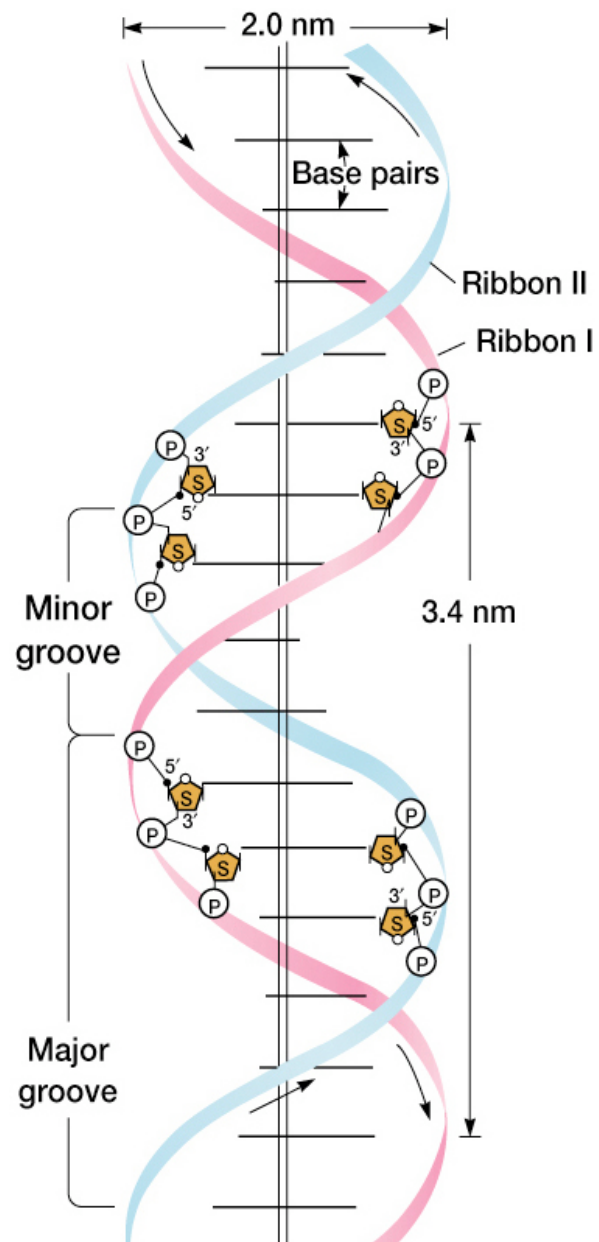
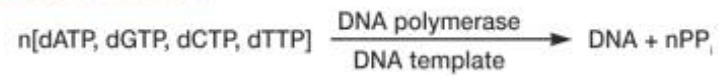


Fig. 11.6

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DNA polymerase reaction



The mechanism of chain growth

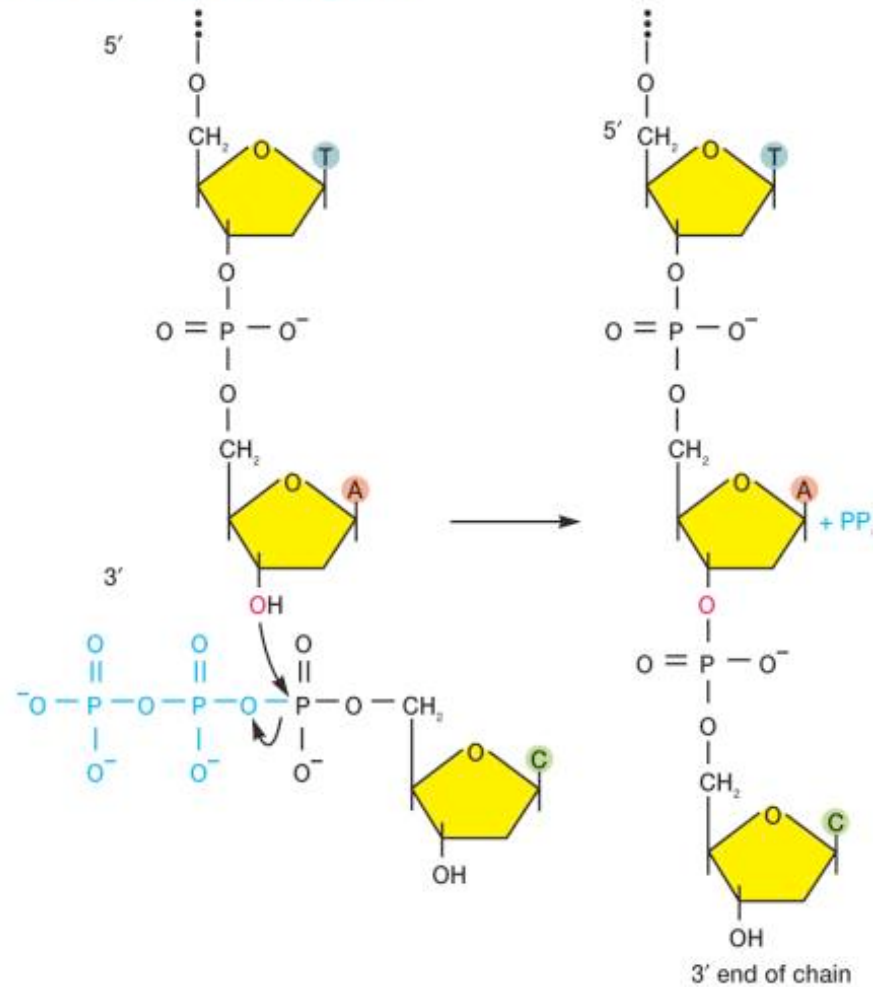


Fig. 11.15

ddATP= stops DNA elongation

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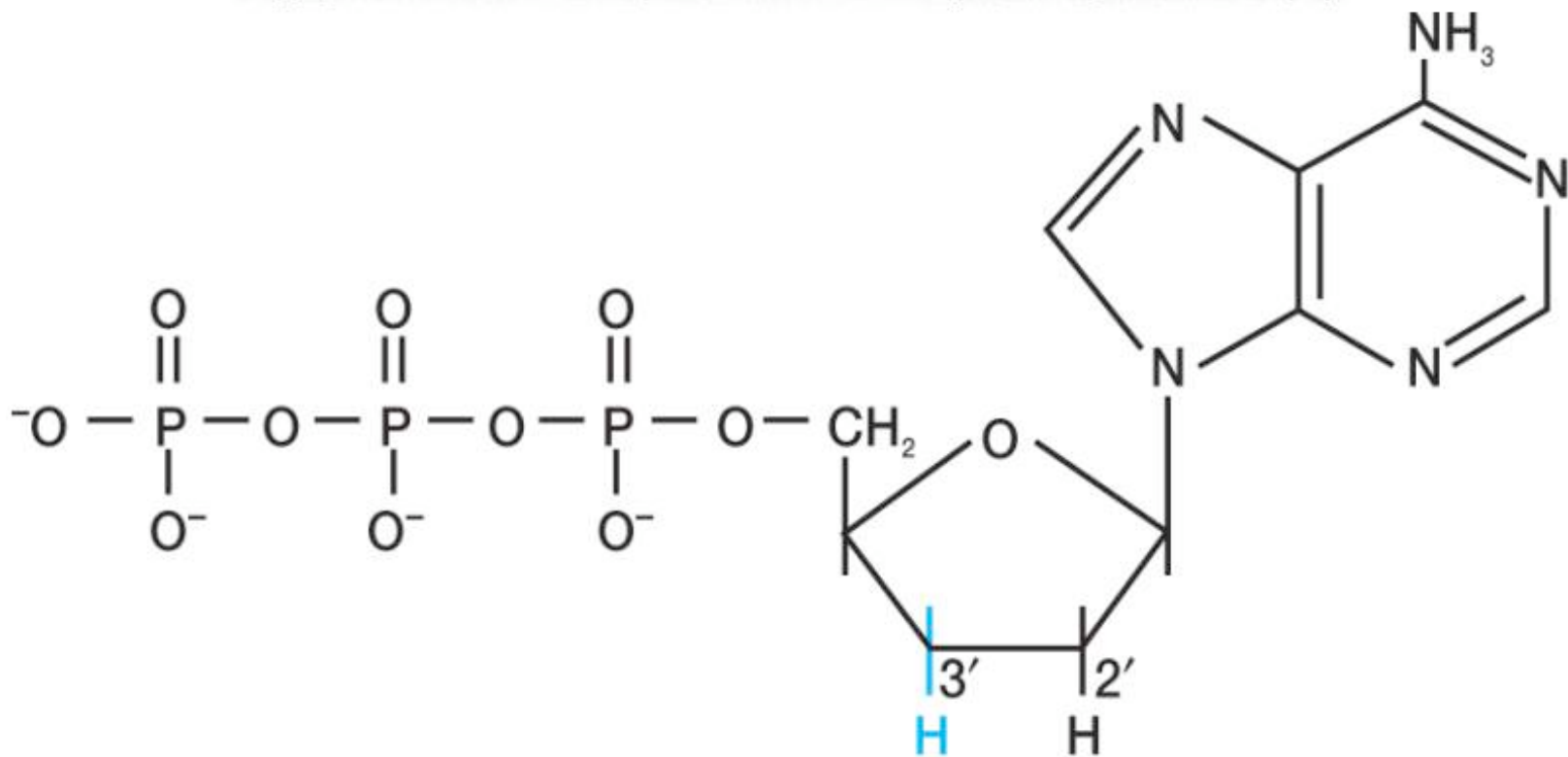
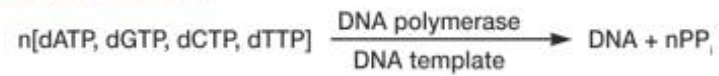


Fig. 15.1

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DNA polymerase reaction



The mechanism of chain growth

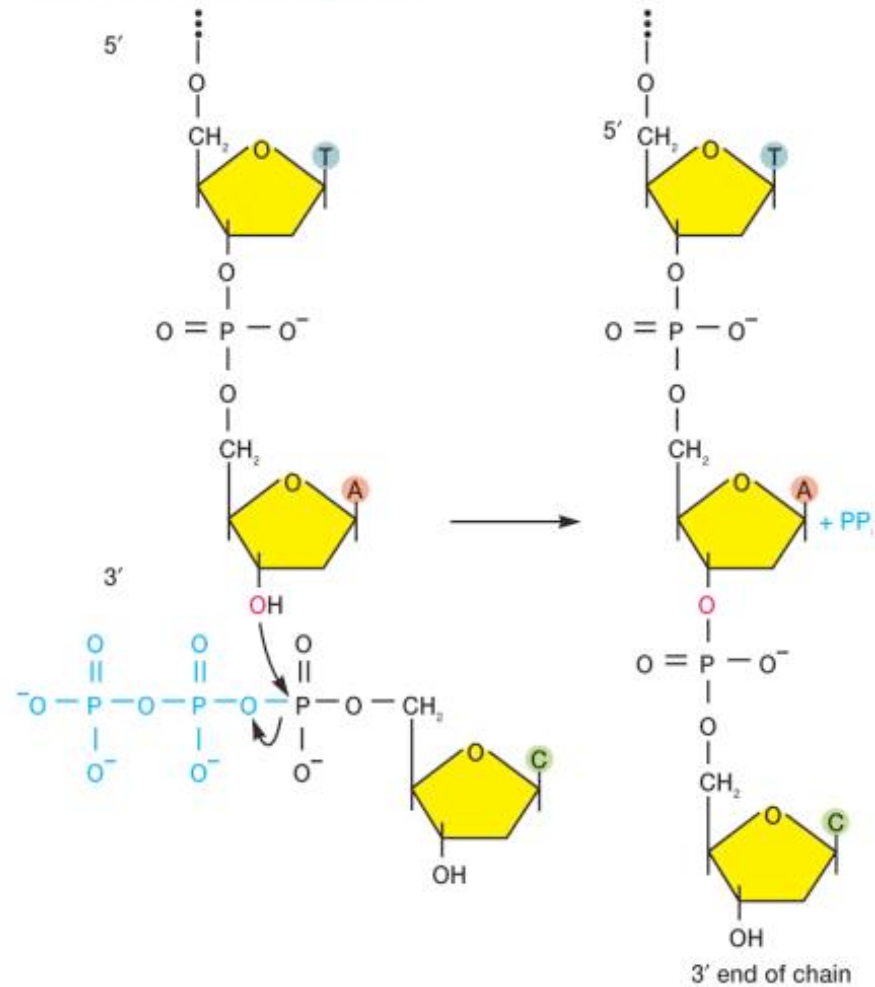


Fig. 11.15

DNA sequencing

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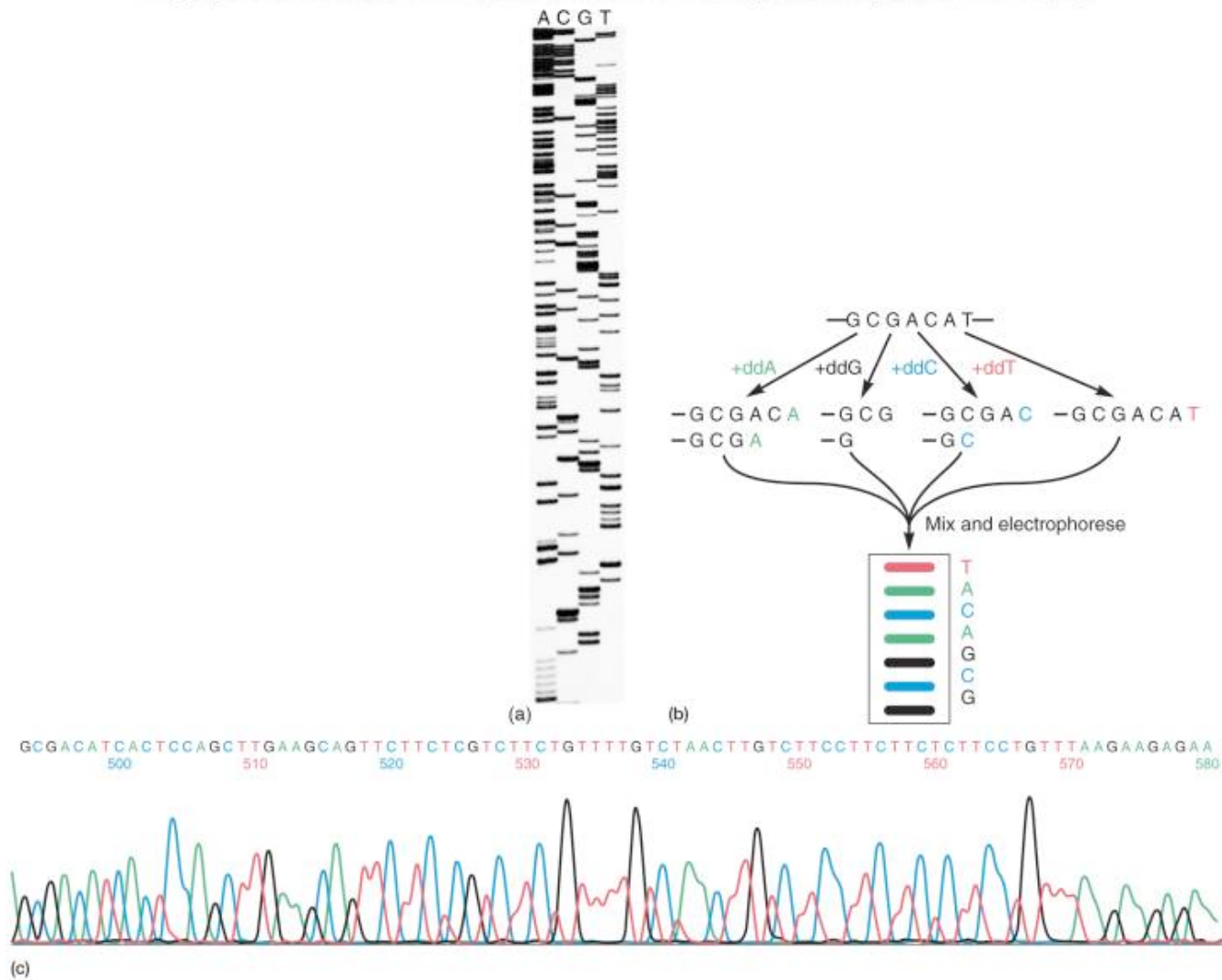


Fig. 15.2

Whole genome sequencing

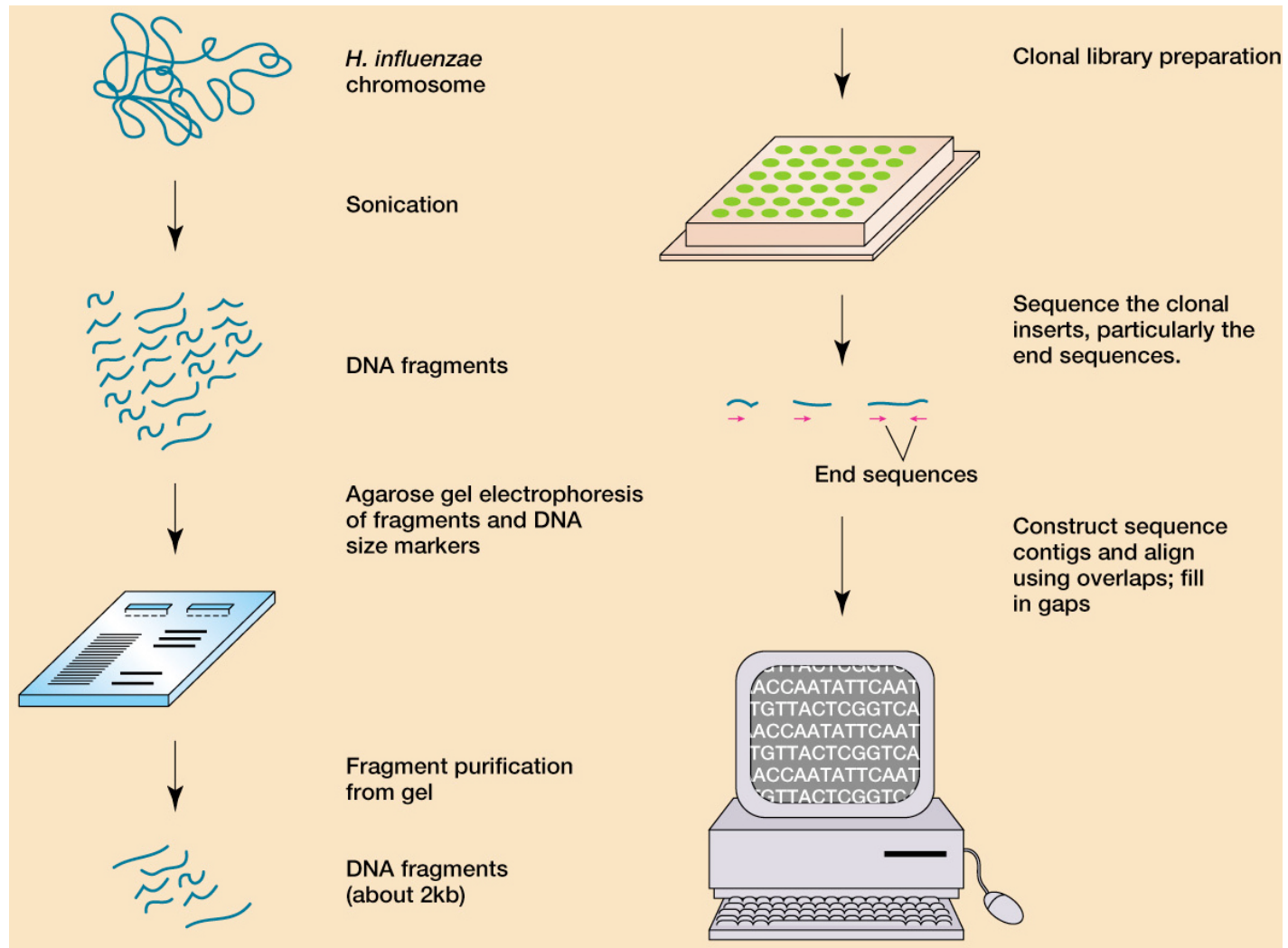


Fig. 15.3

Table 15.1 Examples of Complete Published Microbial Genomes

Genome	Domain ^a	Size (Mb)	% G+C
<i>Aquifex aeolicus</i>	B	1.50	43
<i>Archaeoglobus fulgidus</i>	A	2.18	48
<i>Bacillus subtilis</i>	B	4.20	43
<i>Borrelia burgdorferi</i>	B	1.44	28
<i>Campylobacter jejuni</i>	B	1.64	31
<i>Chlamydia pneumoniae</i>	B	1.23	40
<i>Chlamydia trachomatis</i>	B	1.05	41
<i>Deinococcus radiodurans</i>	B	3.28	67
<i>Escherichia coli</i>	B	4.60	50
<i>Haemophilus influenzae</i> Rd	B	1.83	39
<i>Helicobacter pylori</i>	B	1.66	39
<i>Methanobacterium thermoautotrophicum</i>	A	1.75	49
<i>Methanococcus jannaschii</i>	A	1.66	31
<i>Mycobacterium tuberculosis</i>	B	4.40	65
<i>Mycoplasma genitalium</i>	B	0.58	31
<i>Mycoplasma pneumoniae</i>	B	0.81	40
<i>Neisseria meningitidis</i>	B	2.27	51
<i>Pseudomonas aeruginosa</i>	B	6.3	67
<i>Pyrococcus horikoshii</i>	A	1.80	42
<i>Rickettsia prowazekii</i>	B	1.10	29
<i>Saccharomyces cerevisiae</i>	E	13	38
<i>Synechocystis</i> sp.	B	3.57	47
<i>Thermotoga maritima</i>	B	1.80	46
<i>Treponema pallidum</i>	B	1.14	52
<i>Vibrio cholerae</i>	B	4.0	48

^aThe following abbreviations are used: A, Archaea; B, Bacteria; E, Eucarya.

**69 Complete Microbial Genomes
(as of 12/17/01)**

[A] - Archaea - 12 species

[B] - Bacteria - 57 species

**224 Complete Microbial Genomes
(as of 2/17/05)**

[A] - Archaea - 21 species

[B] - Bacteria - 203 species

Map of *Mycoplasma genitalium*

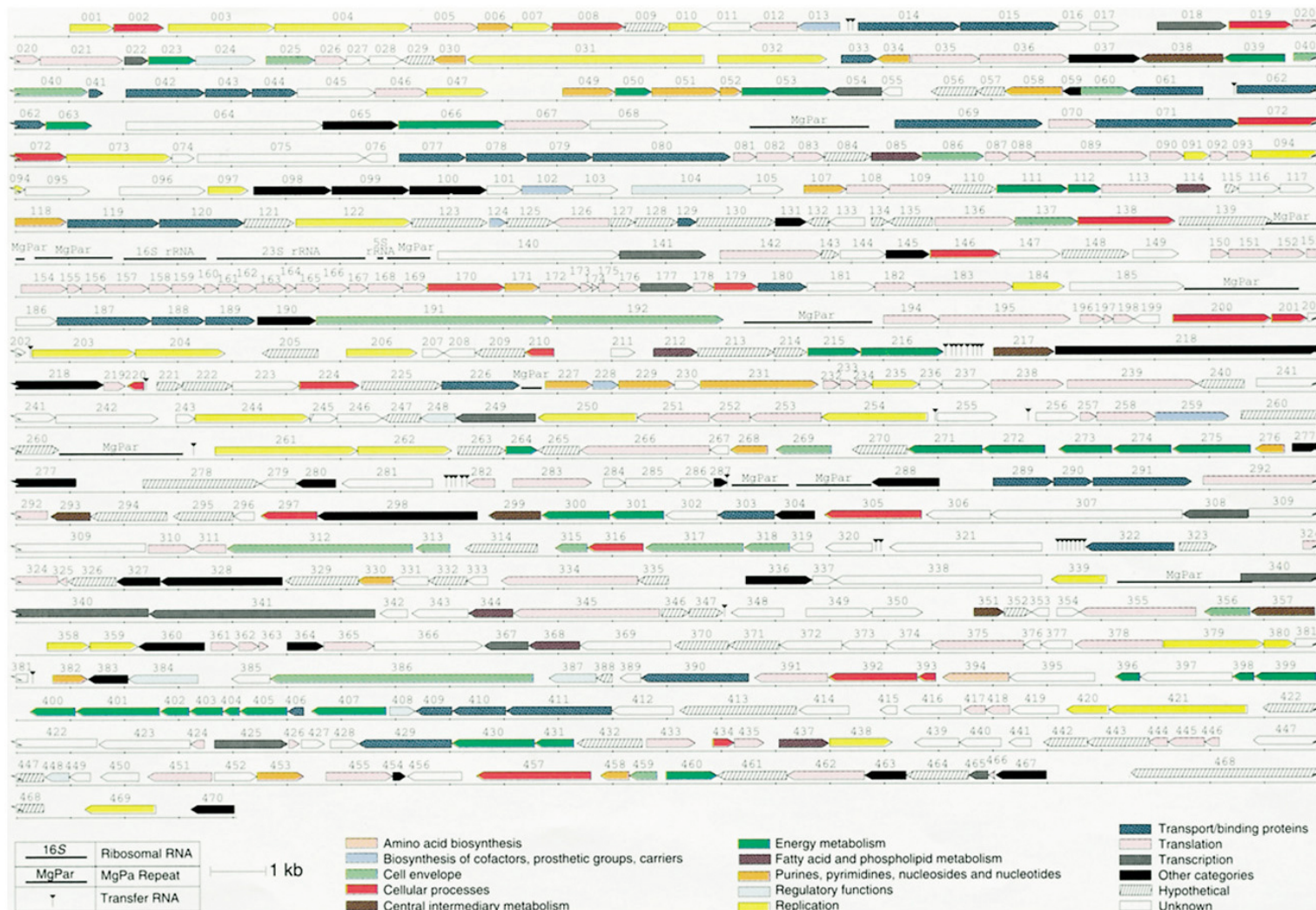
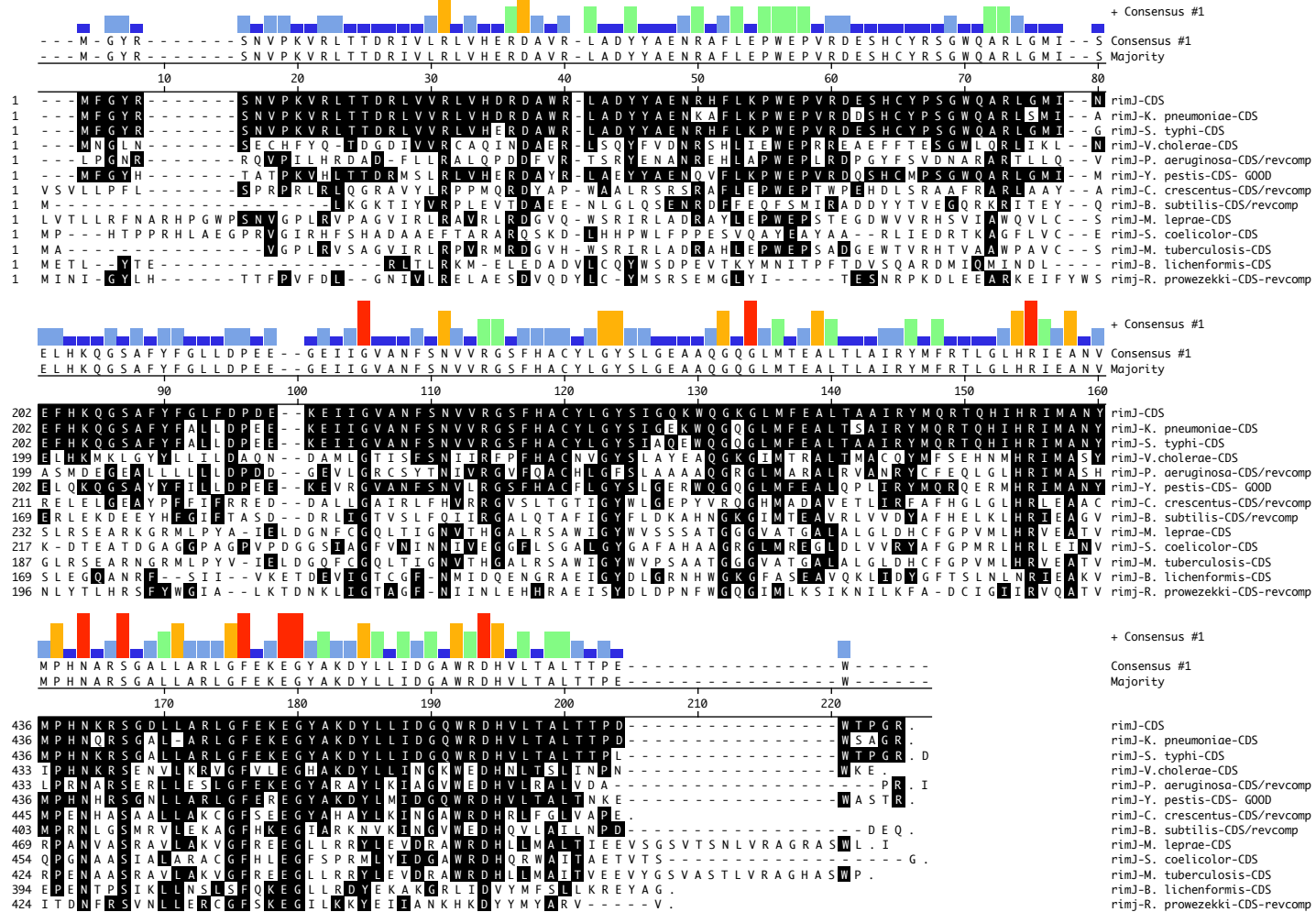


Fig. 15.5

What do you get from sequence data?

- Evolutionary relatedness based on several loci
- Infer function of proteins encoded by genes

Multiple sequence alignment



The RimJ protein

MFGYRSNVPKVRLTTDRLVVRRLVHDRDAWRLADYYAENRHFLKPWEPVRD

Motif C

↓
L

ESHYCPSGWQARLGMINFHKQGSIFYFGLFDPDEKEIIGVANFSNVVRG

Motif D

↓
stop

SFHACYLGYSIGQKWQGKGLMFEALTAAIRYMORTQHIHRIMANYMPHNK

Motif A

↑
rimJ-2::mTn10

RSGDLLARLGFEKEEGYAKDYLLIDGQWRDHVLTAALTTPDWTPGR.

Motif B

↑
rimJ-1::mTn10

Red= conserved in RimJ and 13 putative prokaryotic acetylases

Yellow= conserved in RimJ and 11 putative prokaryotic acetylases

Blue= *rimJ* mutations that cause a loss of *papBA* thermoregulation

Green= motifs conserved in N-acetyltransferase (GNAT) superfamily

Map of *Haemophilus influenzae*

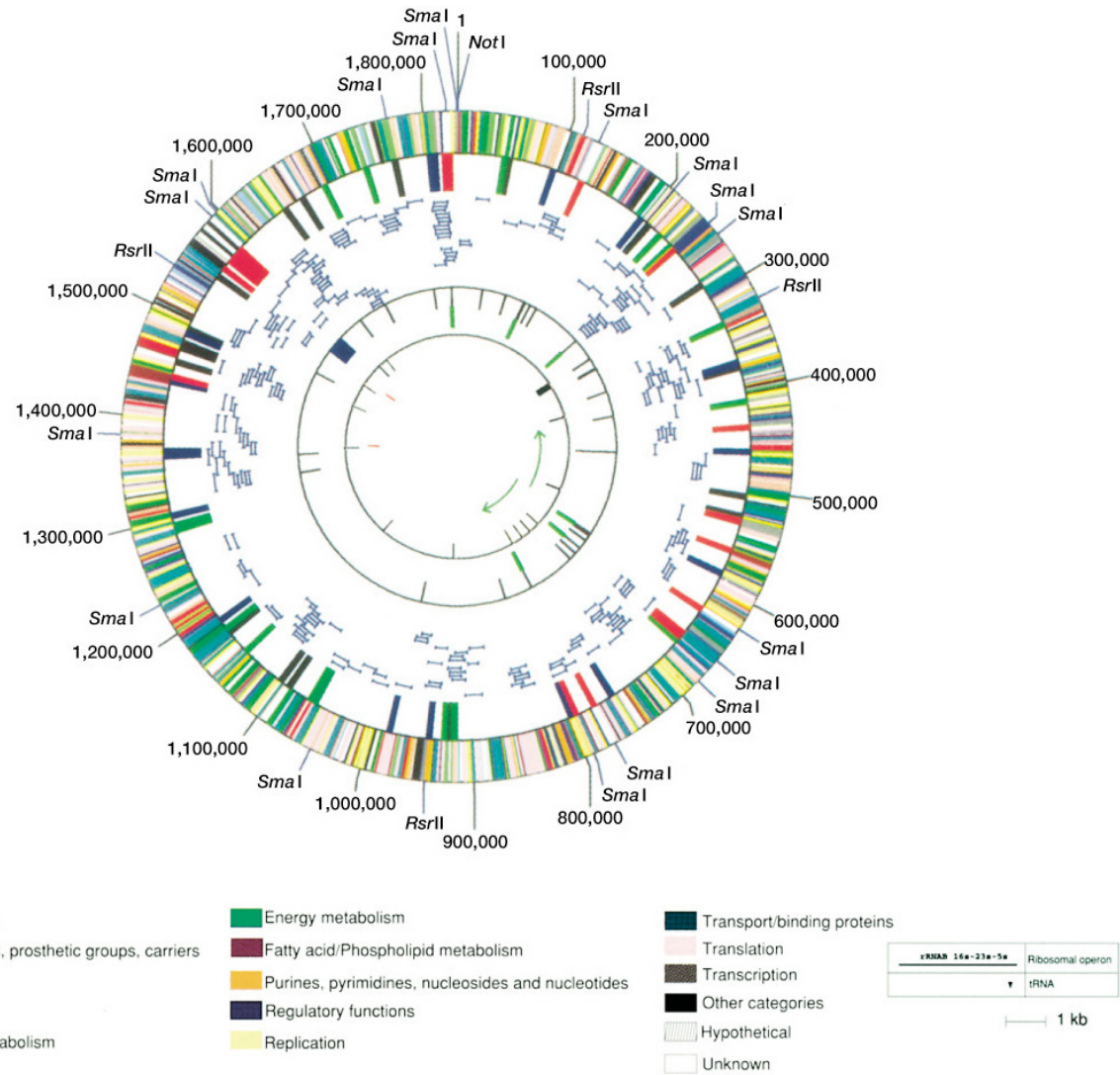
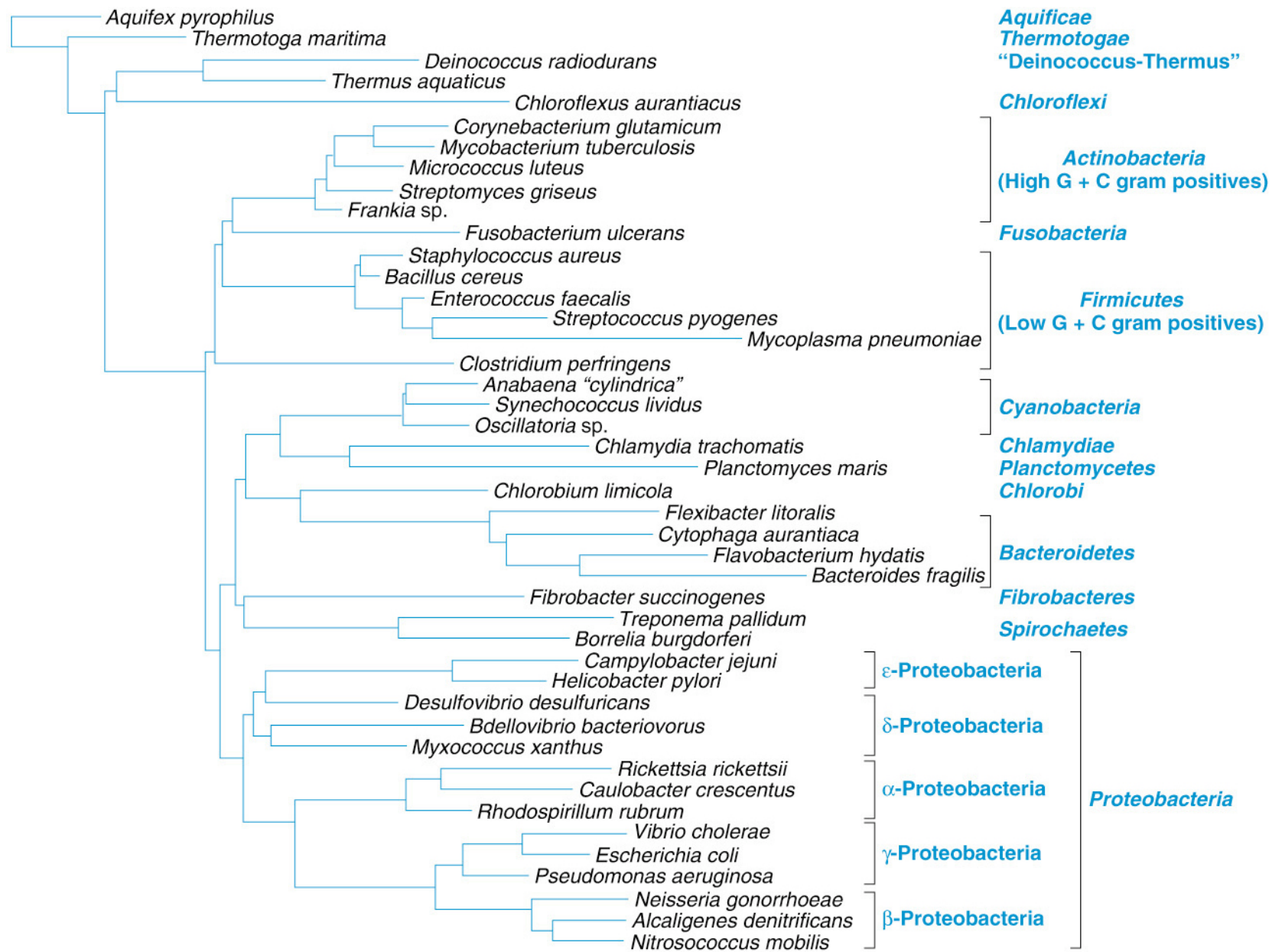


Fig.15.6



Bergey's Manuals

- *Manual of Determinative Bacteriology*
 - used for identification of bacterial species
- *Manual of Systematic Bacteriology*
 - more thorough taxonomic classification
 - 1st edition= phenetic classification
 - 2nd edition= phylogenetic classification